

# Small Angle X-ray Scattering

## Overview of basic data evaluation and analysis

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Biomolekulare NMR-Spektroskopie

## ATSAS Package

Collection of tools for data evaluation  
and modeling of SAXS and SANS data

EMBL



<https://www.embl-hamburg.de/biosaxs/software.html>

### Platforms:

Linux

Windows

Mac OS

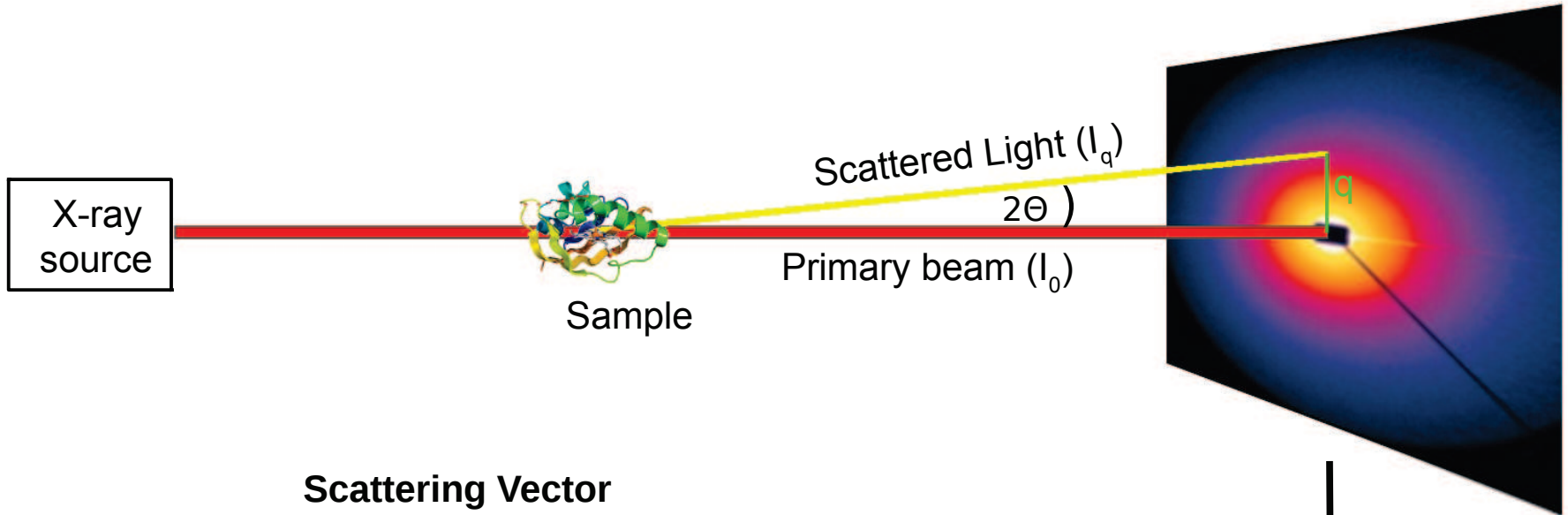
For academic use the software is free.

### Download:

<https://www.embl-hamburg.de/biosaxs/download.html>

(Registration required)

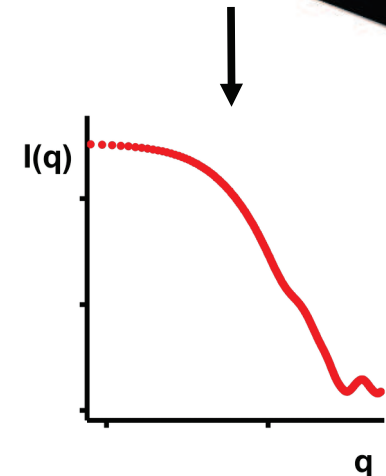
## Small Angle X-ray Scattering Experiment



### Scattering Vector

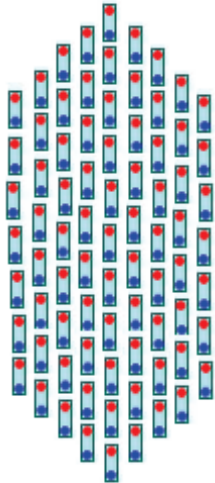
$$|\vec{q}| = \frac{4\pi}{\lambda} \sin 2\Theta \quad \vec{q} = (\vec{k}_s - \vec{k}_0)$$

$k_0, k_s$  Wave vectors of incident and scattered beam  
 $\lambda$  Wavelength of incident beam

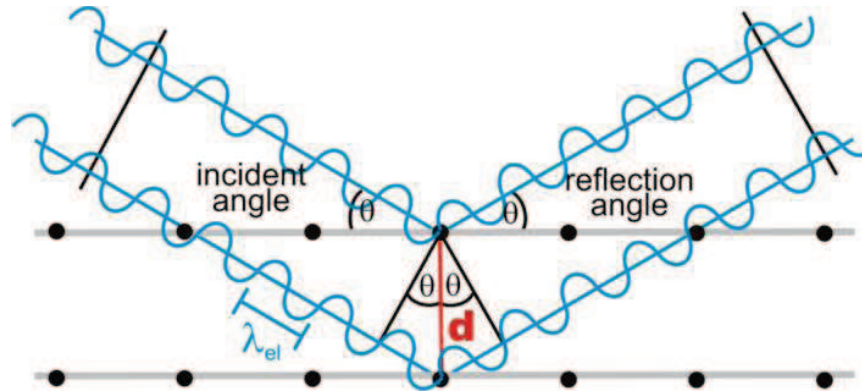


## Small angle scattering and Single crystal diffraction

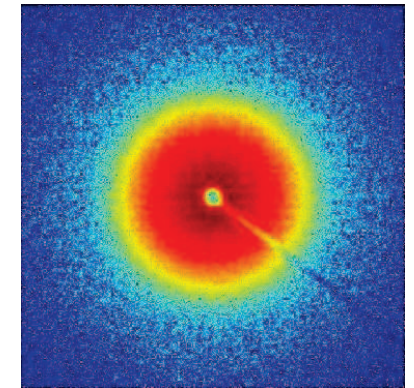
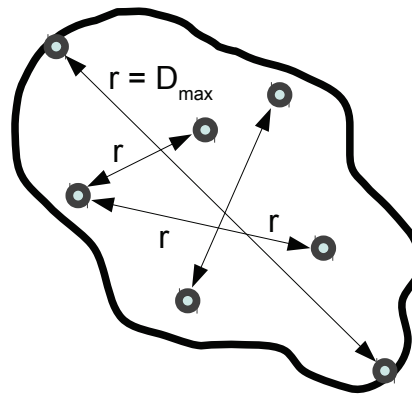
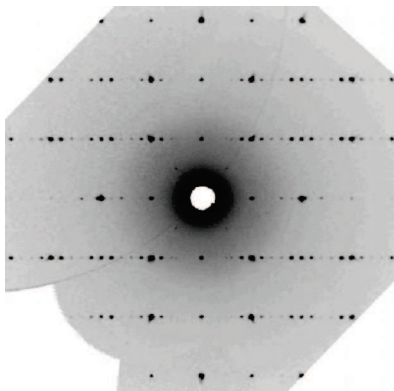
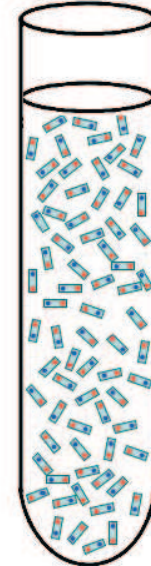
Defined orientation of molecules in a crystal

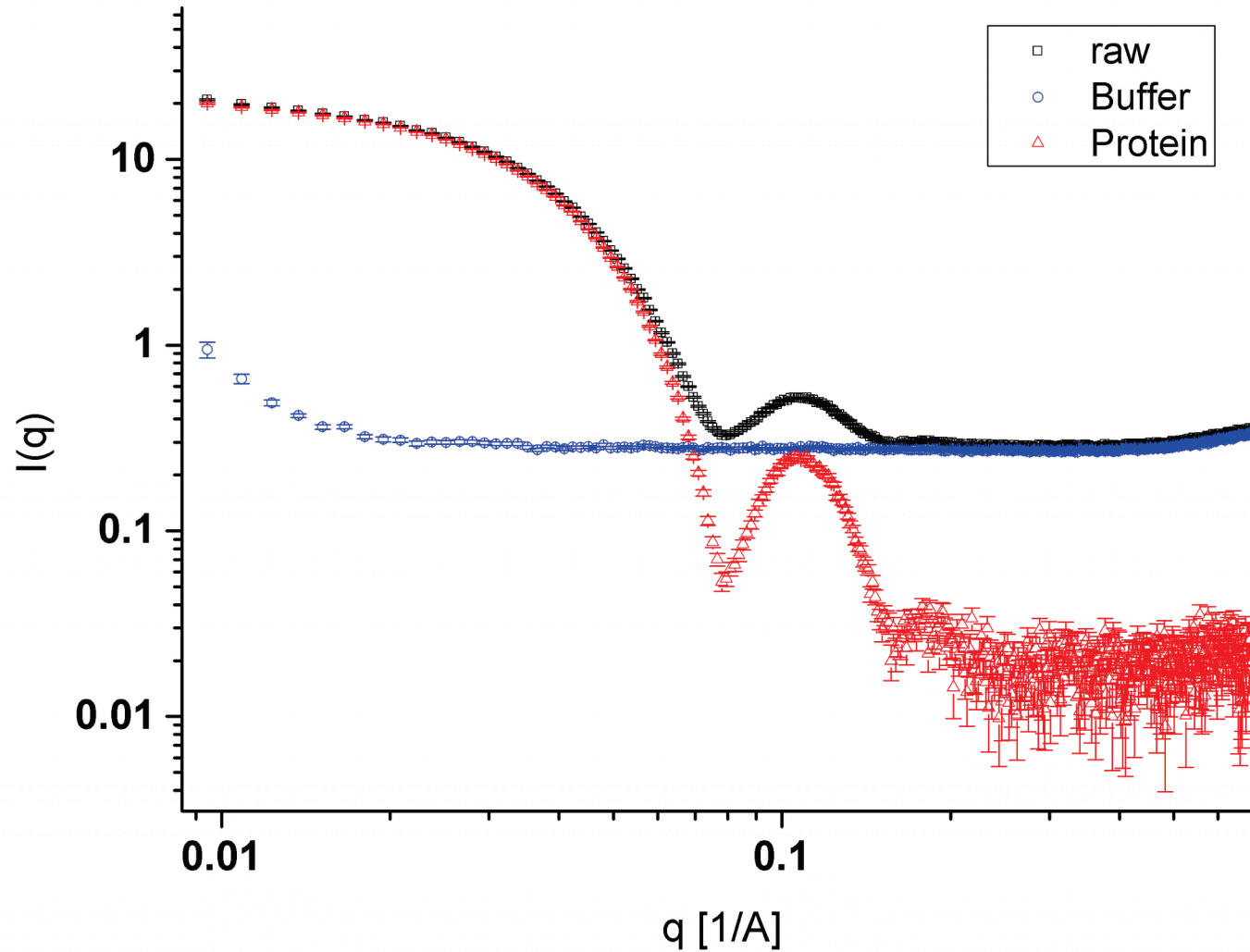


Braggs Law



Random orientation of molecules in solution





## What can we learn from Small Angle X-ray Scattering Curves

**What we put in = what we get out**

## What can we learn from Small Angle X-ray Scattering Curves

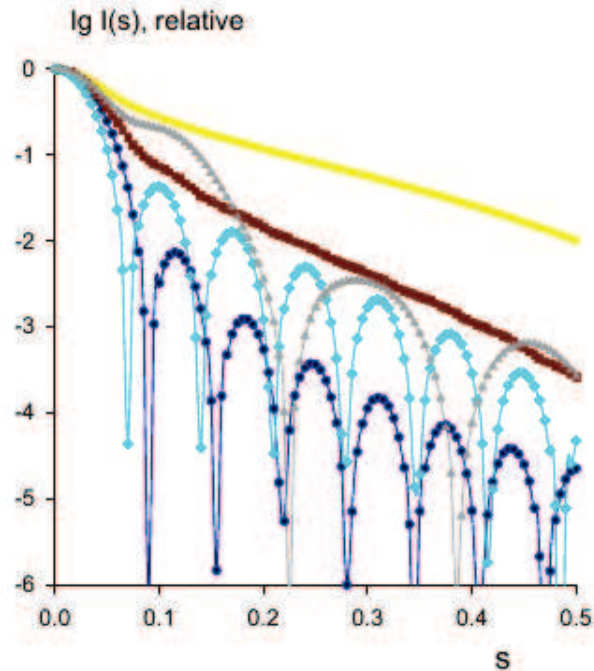
**Garbage in = Garbage out**

## What can we learn from Small Angle X-ray Scattering Curves

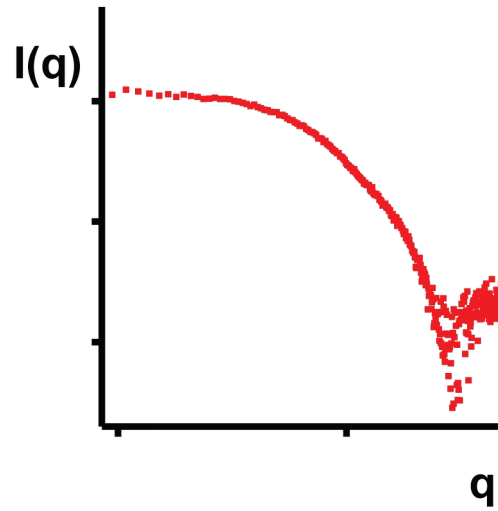
**More secondary information in  
=  
Much more information out**

# Direct Information

## Shape

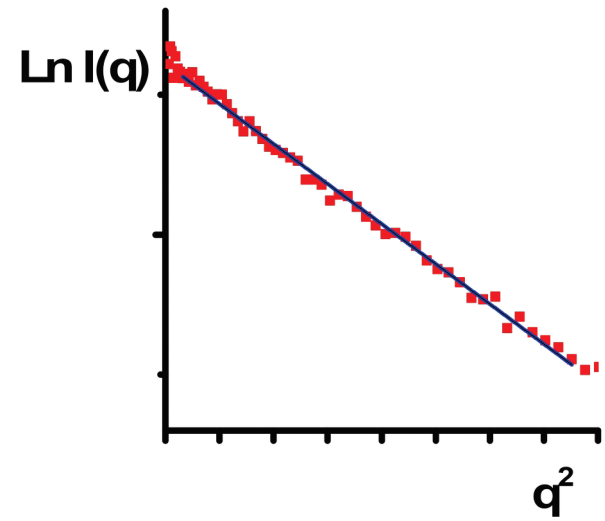


Scattering curve



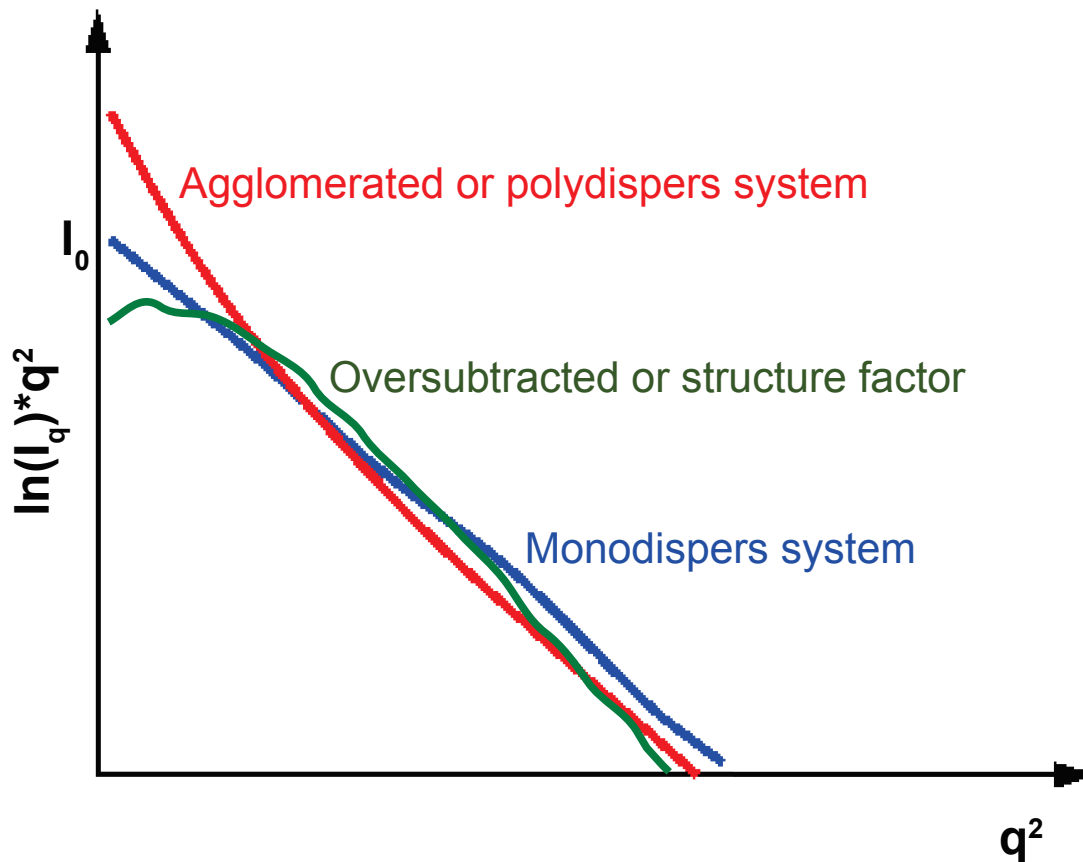
- Radius of Gyration
- $I_0$  and Molecular Weight
- Dimerisation or ligand binding

Guinier Plot



## Guinier plot

**Radius of Gyration ( $R_g$ ):** Average of square centers of mass distances in a particle



Objects of same mass



### Guinier Approximation

$$I(q) \approx I_0 e^{-\frac{R_g^2}{3} q^2}$$

linearisation

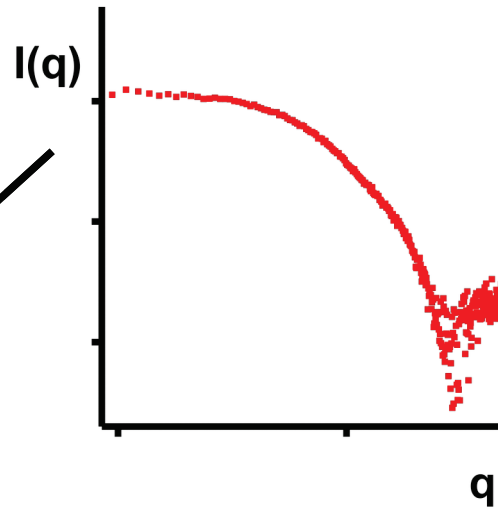
$$\ln(I_q) = -\frac{1}{3} R_g^2 q^2 + I_0$$

Limit:  $R_g * q < 1.3$

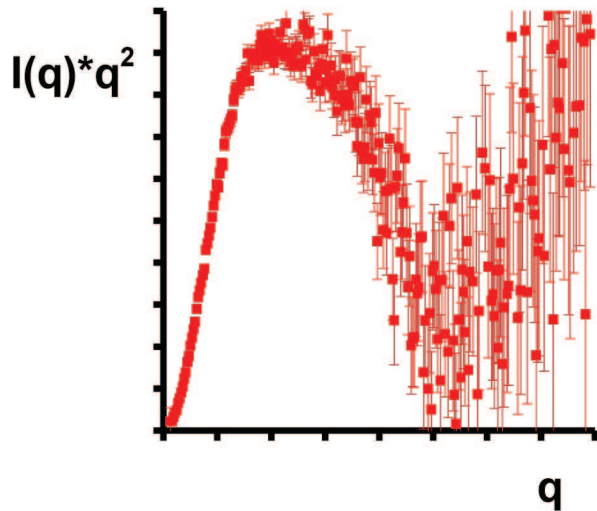
Scattering curve

- State of folding  
folded  
unfolded  
partially folded

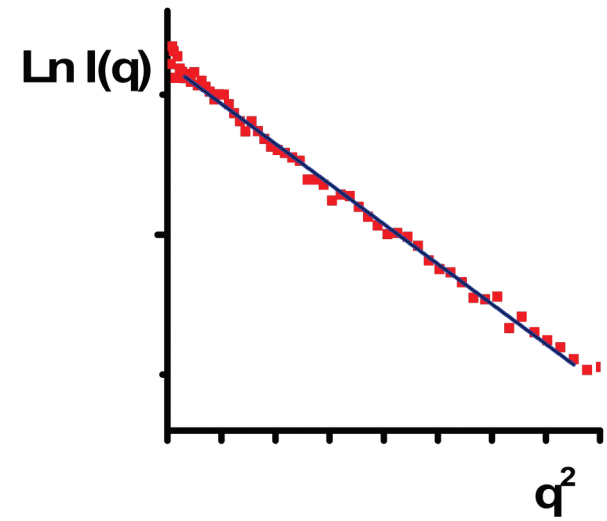
- Radius of Gyration
- $I_0$  and Molecular Weight
- Dimerisation or ligand binding



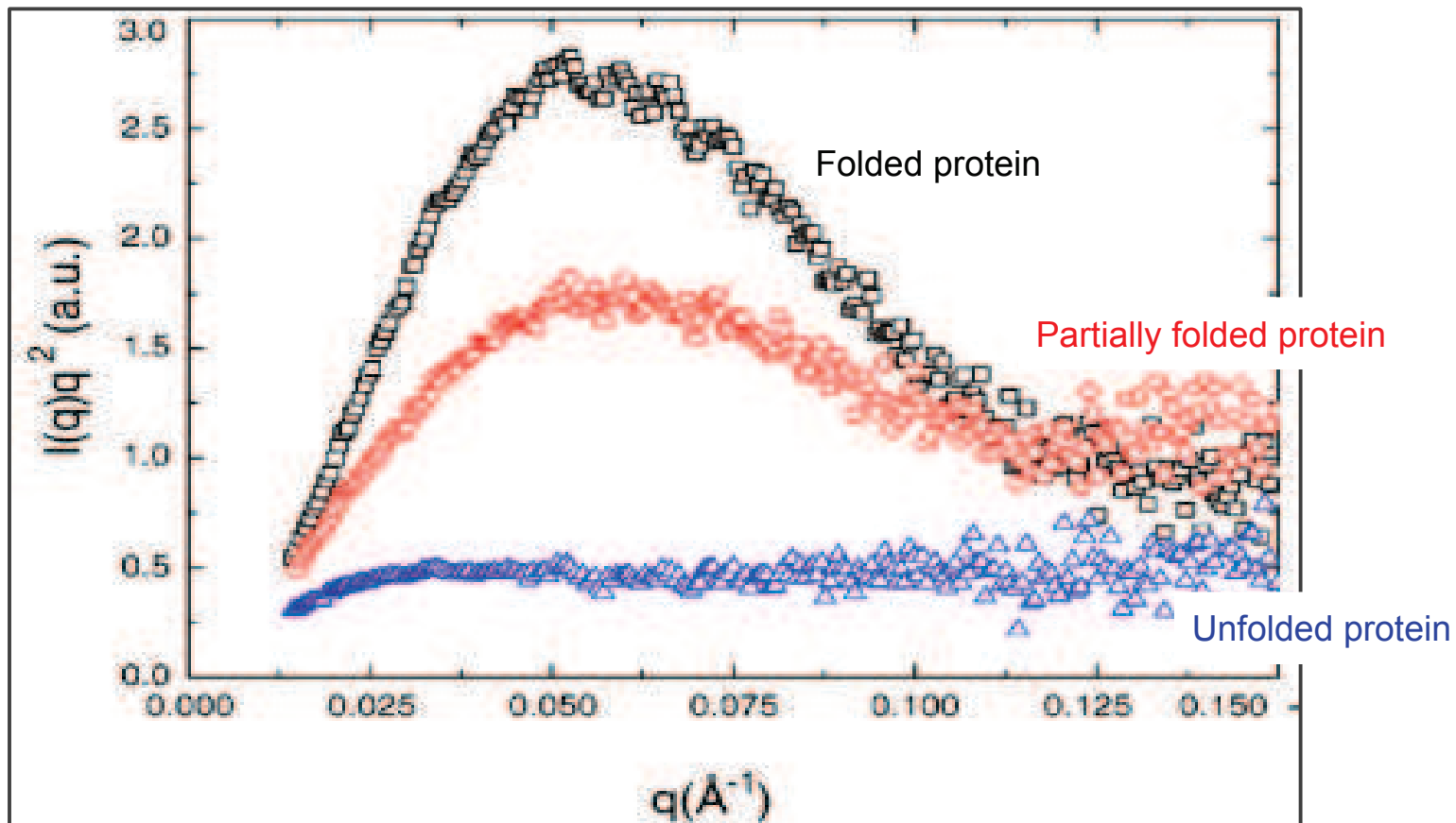
Kratky Plot



Guinier Plot



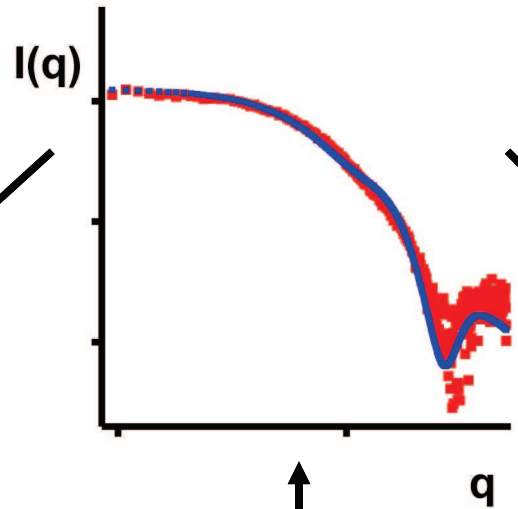
## Kratky plot



The Kratky plot shows the state of folding of a protein (BSA with increasing concentrations of urea)

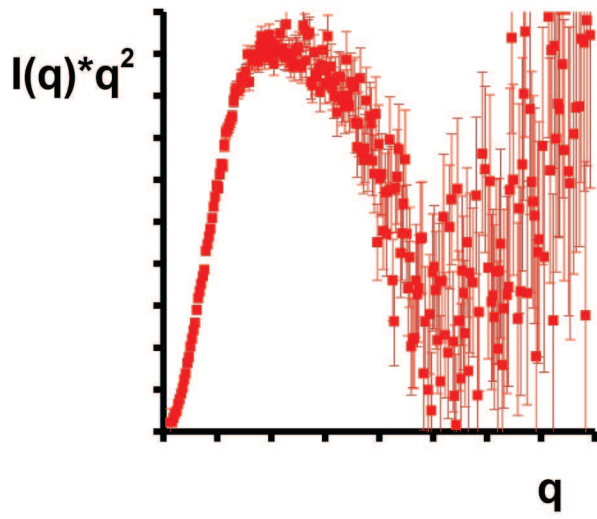
- State of folding  
folded  
unfolded  
partially folded

Fit of crystal structure  
to SAXS data

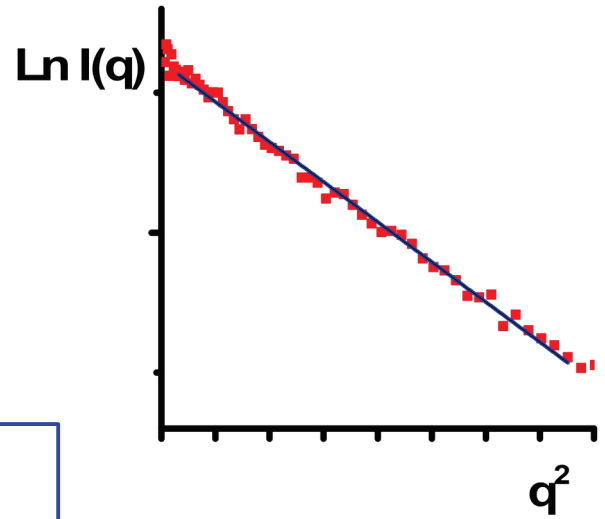


- Radius of Gyration
- $I_0$  and Molecular Weight
- Dimerisation or ligand binding

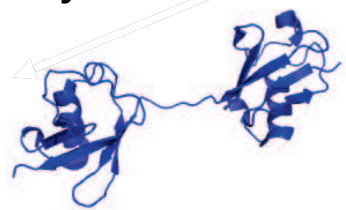
Kratky Plot



Guinier Plot



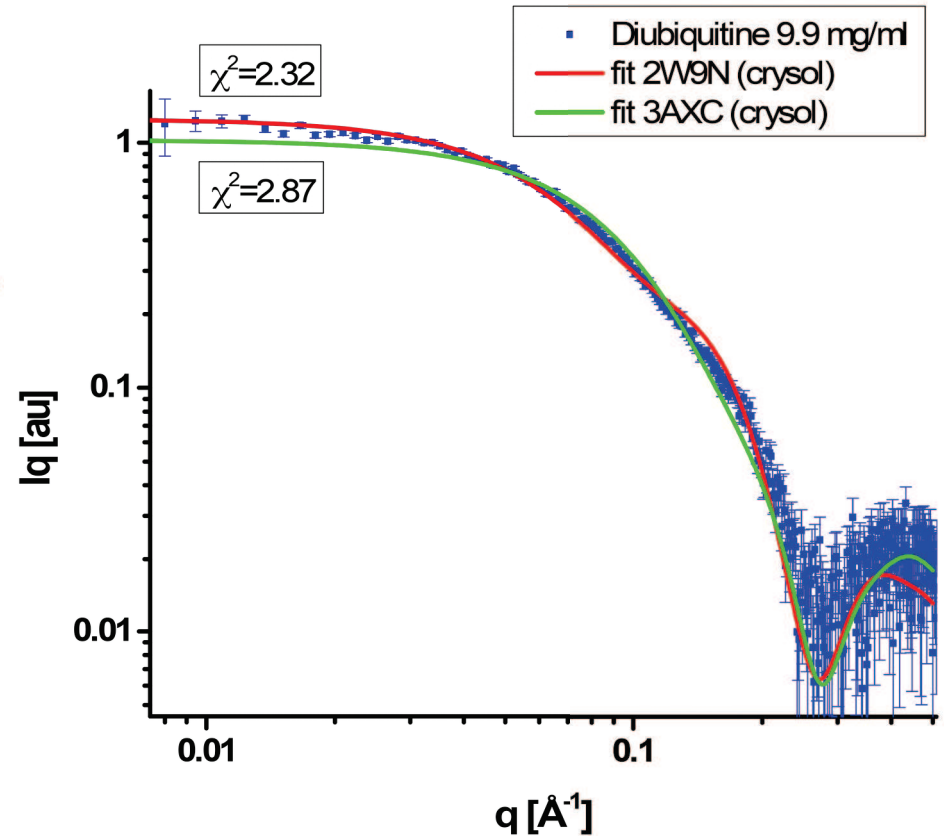
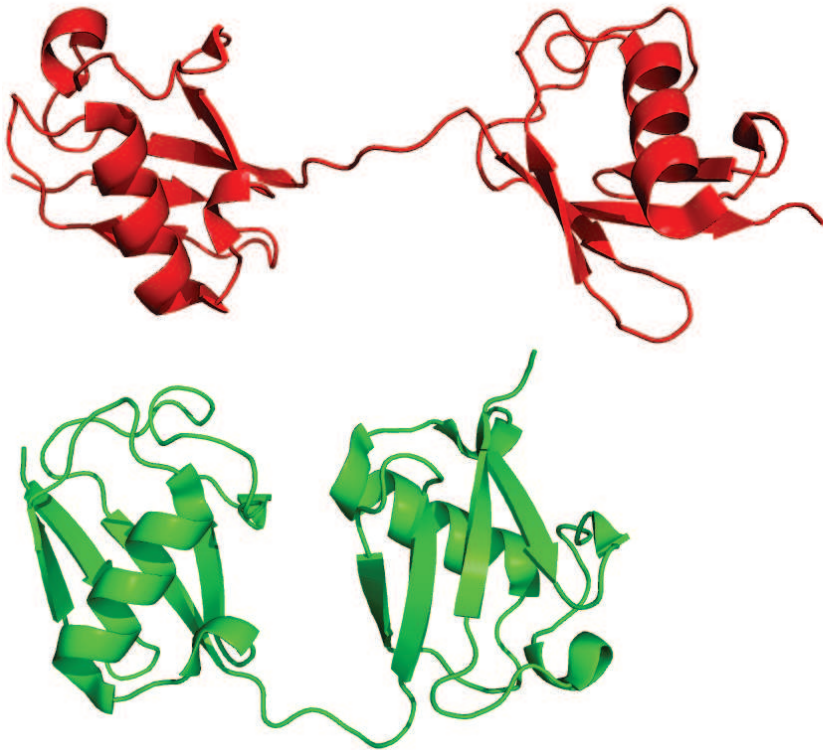
Crystal Structure



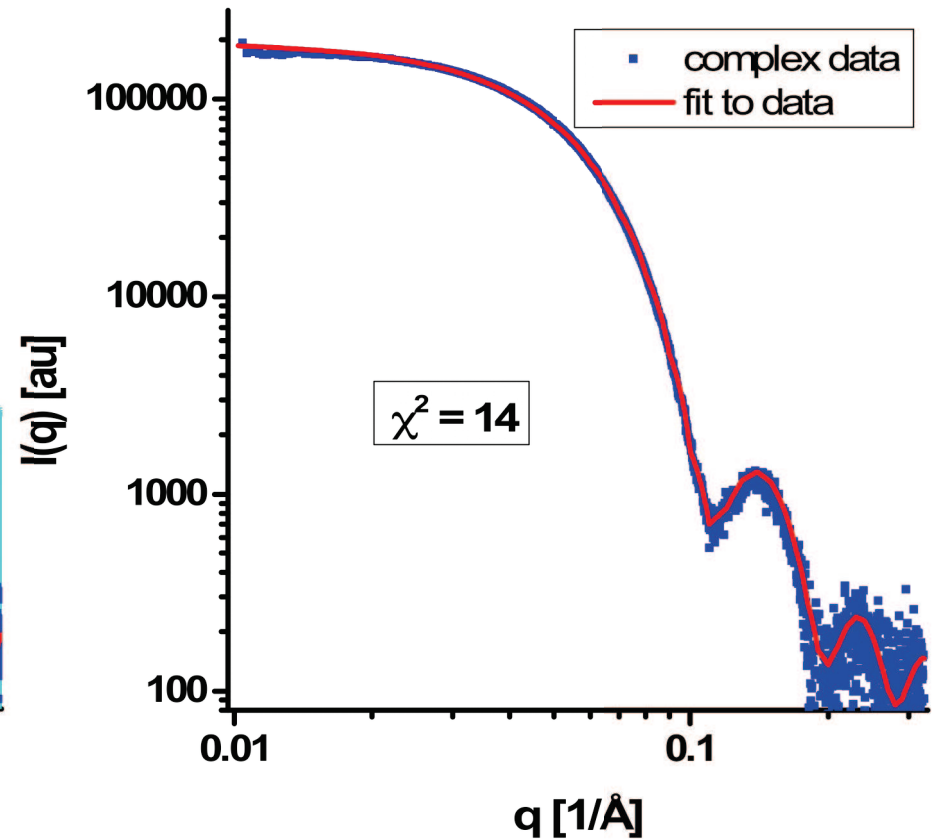
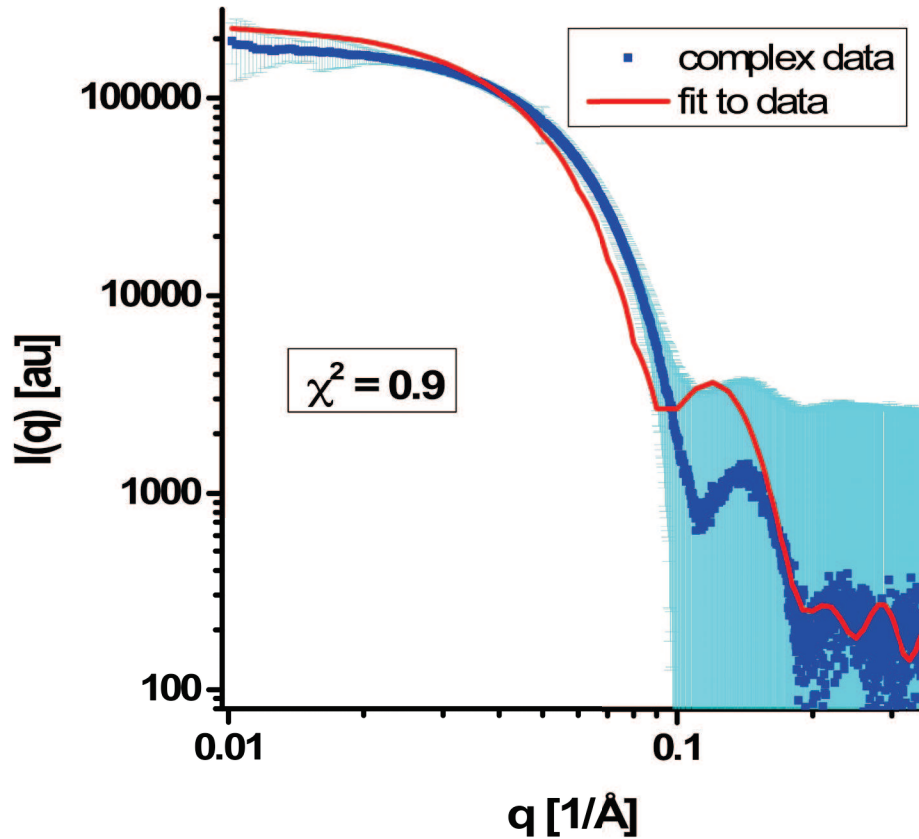
- Difference between crystal structure and structure in solution
- Dimerisation

## Combination of high resolution models with SAXS CRY SOL

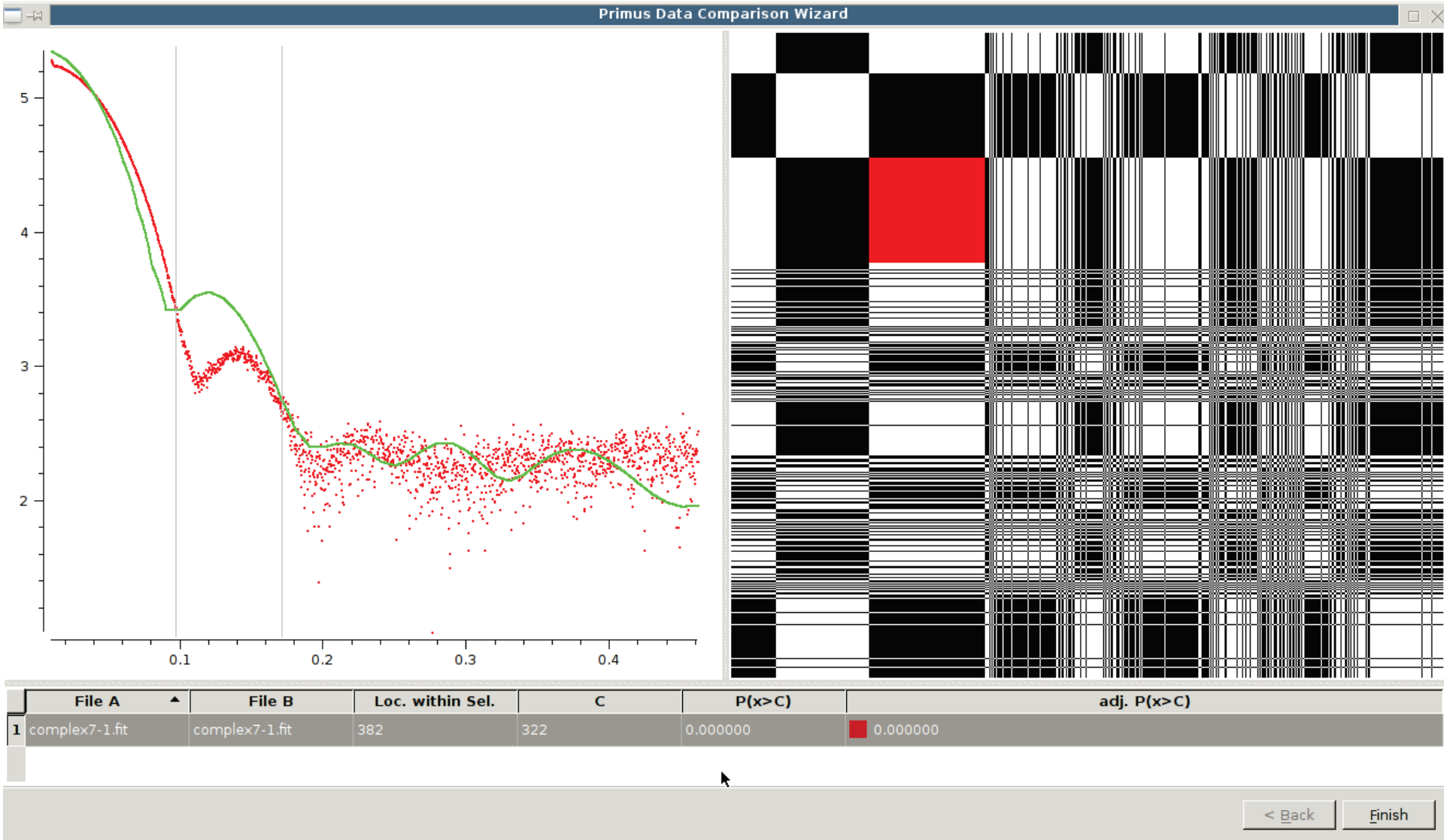
crystal calculates a scattering profile from a high resolution structure and compares to scattering data



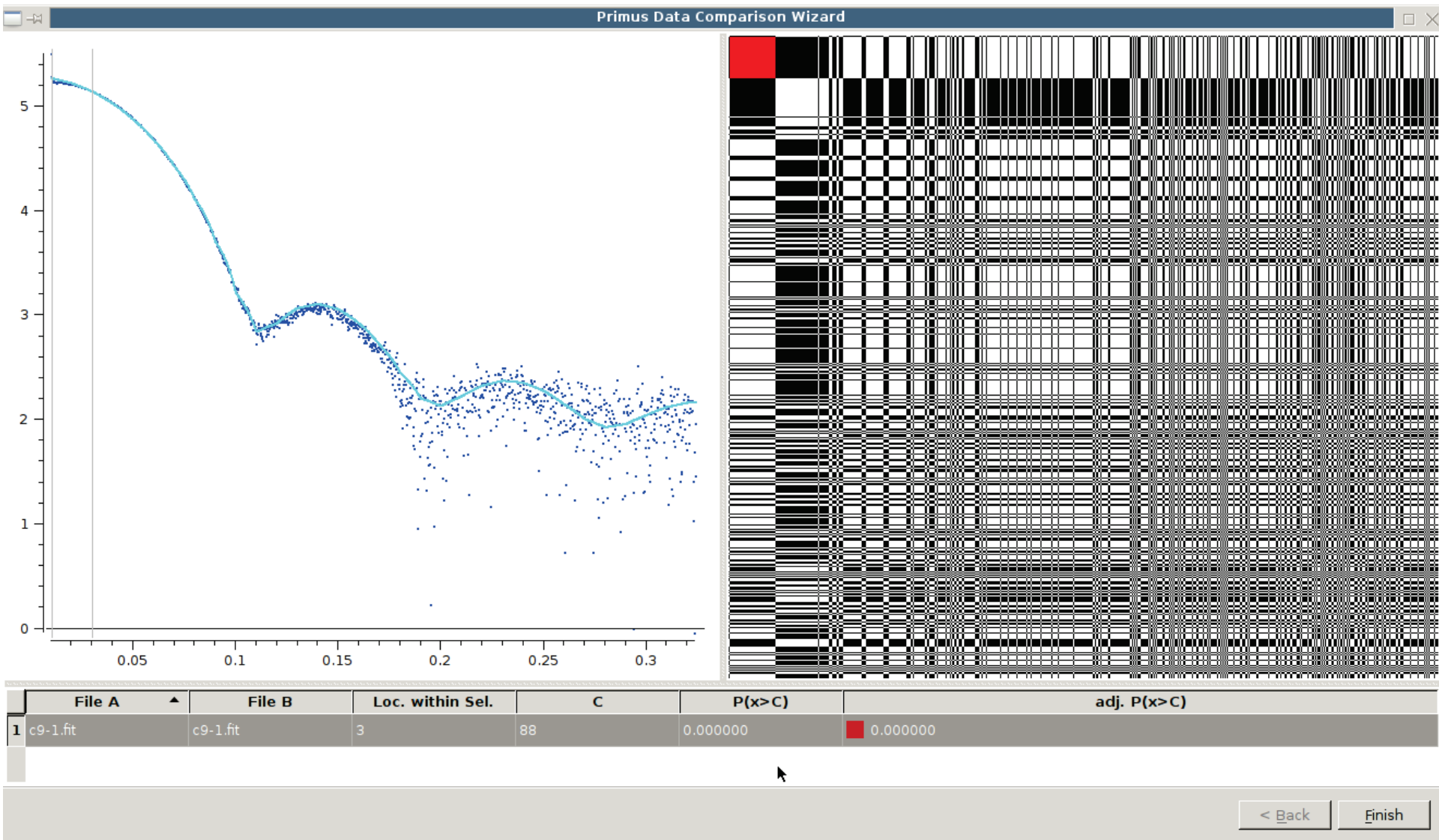
## CRYSOL and $\chi^2$ values



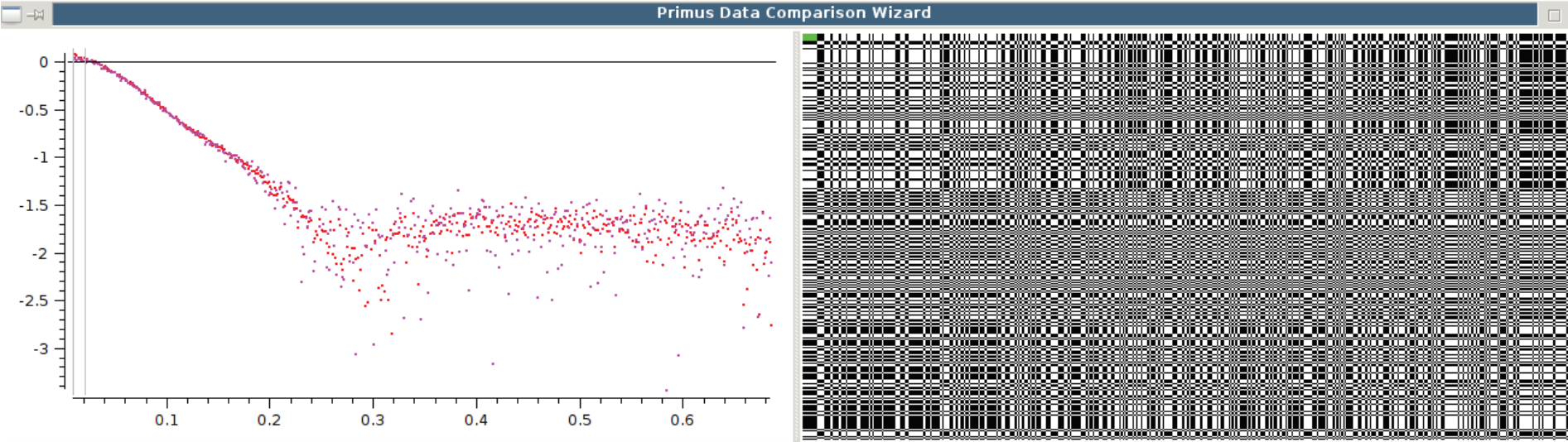
## Correlation map primus



## Correlation map primus



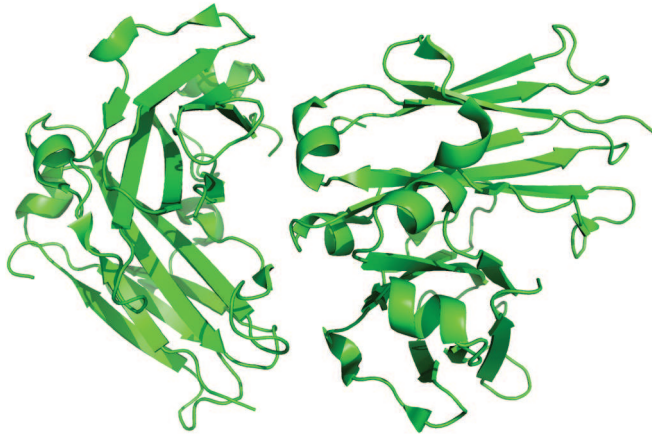
## Correlation map primus



	File A	File B	Loc. within Sel.	C	P(x>C)	adj. P(x>C)
1	diubiquitin_code_9.9...	diubiquitin_code_2.7...	50	8	0.604459	1.000000
2	diubiquitin_code_9.9...	diubiquitin_code_7.7...	57	10	0.204044	1.000000
3	diubiquitin_code_9.9...	diubiquitin_code_5.9...	1	9	0.368285	1.000000
4	diubiquitin_code_2.7...	diubiquitin_code_7.7...	13	6	0.979022	1.000000
5	diubiquitin_code_2.7...	diubiquitin_code_5.9...	390	10	0.204044	1.000000
6	diubiquitin_code_7.7...	diubiquitin_code_5.9...	446	9	0.368285	1.000000

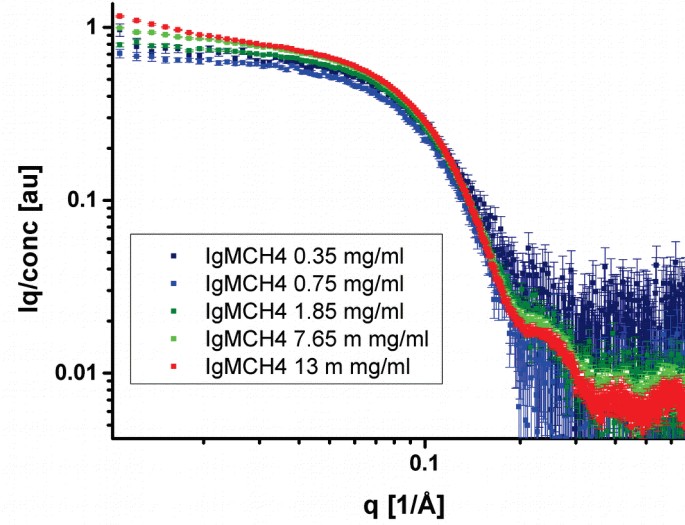
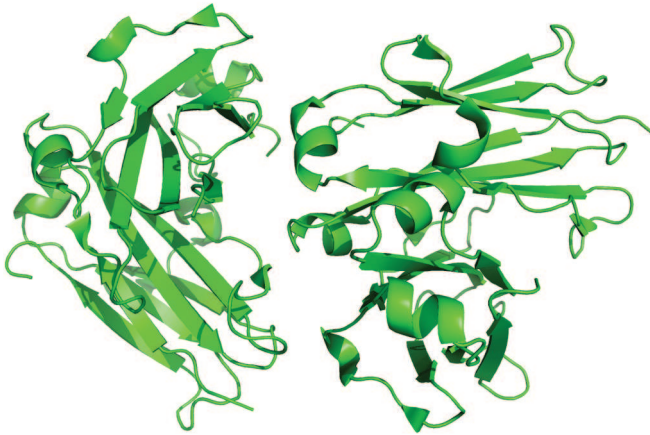
## Dimerisation oligomer

IGMCH4 Dimer



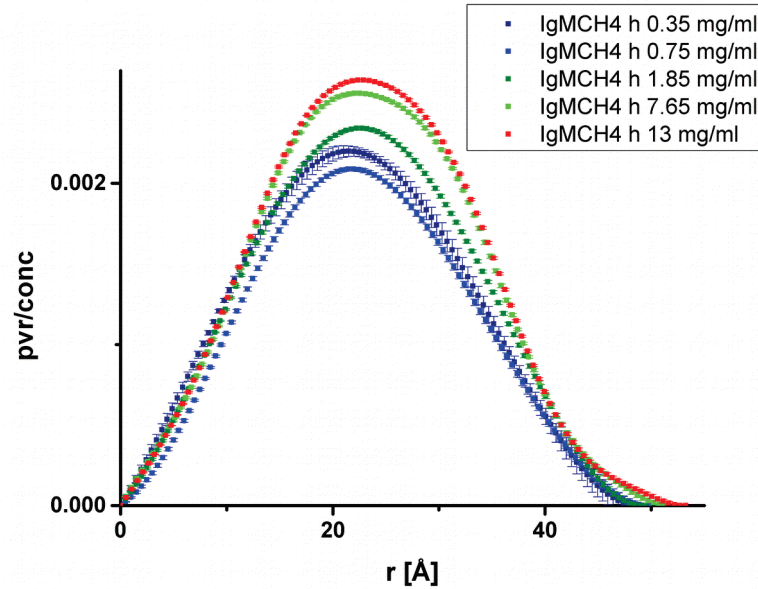
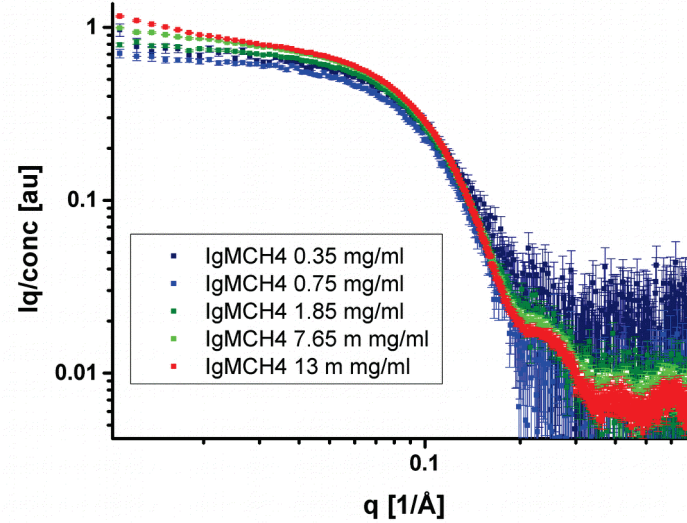
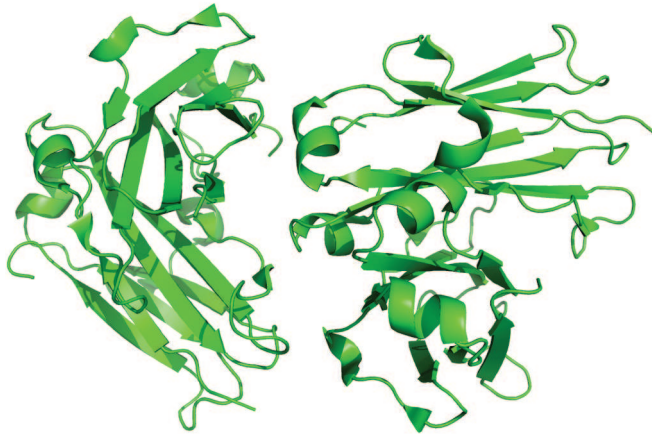
## Dimerisation oligomer

IGMCH4 Dimer



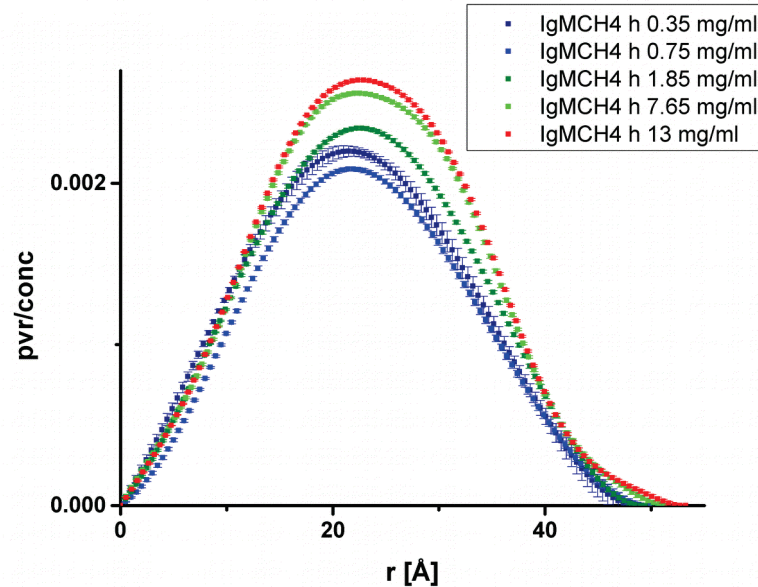
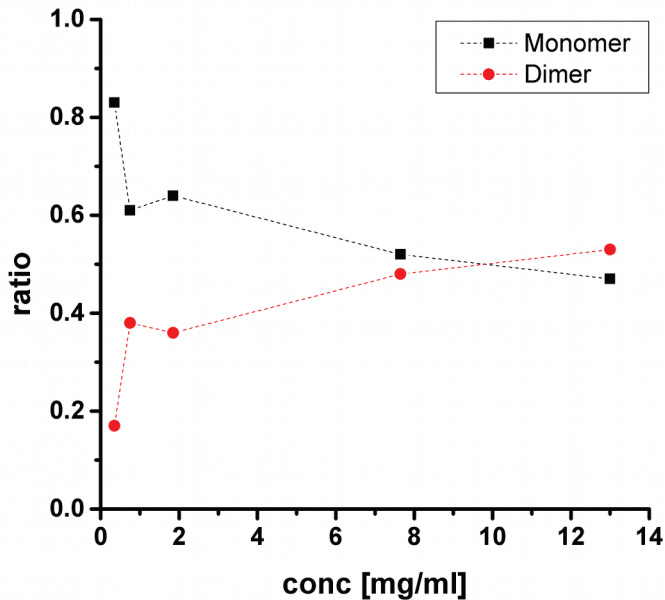
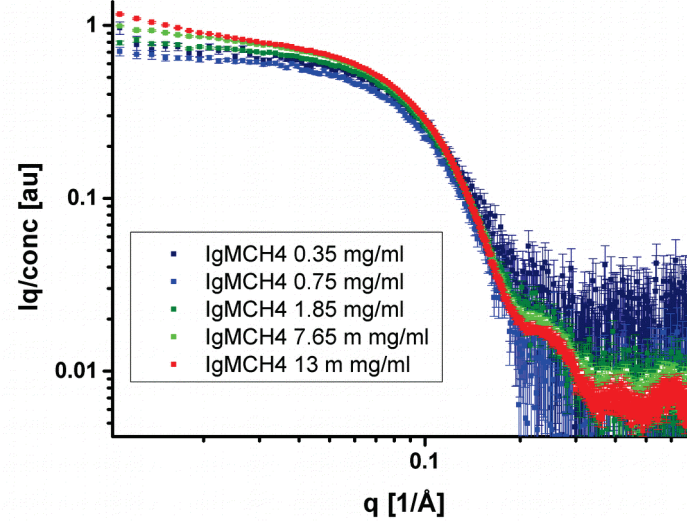
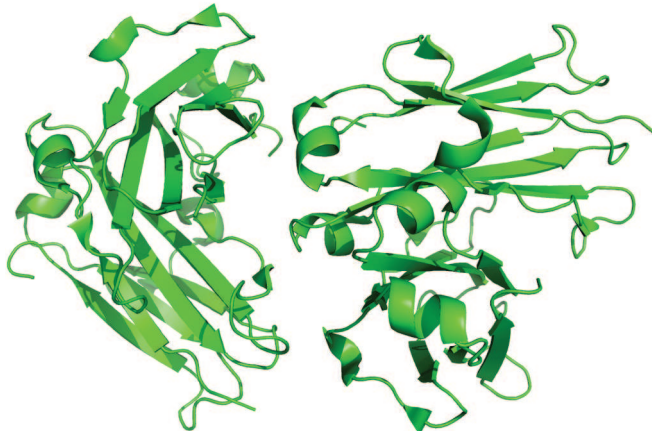
## Dimerisation oligomer

IGMCH4 Dimer



## Dimerisation oligomer

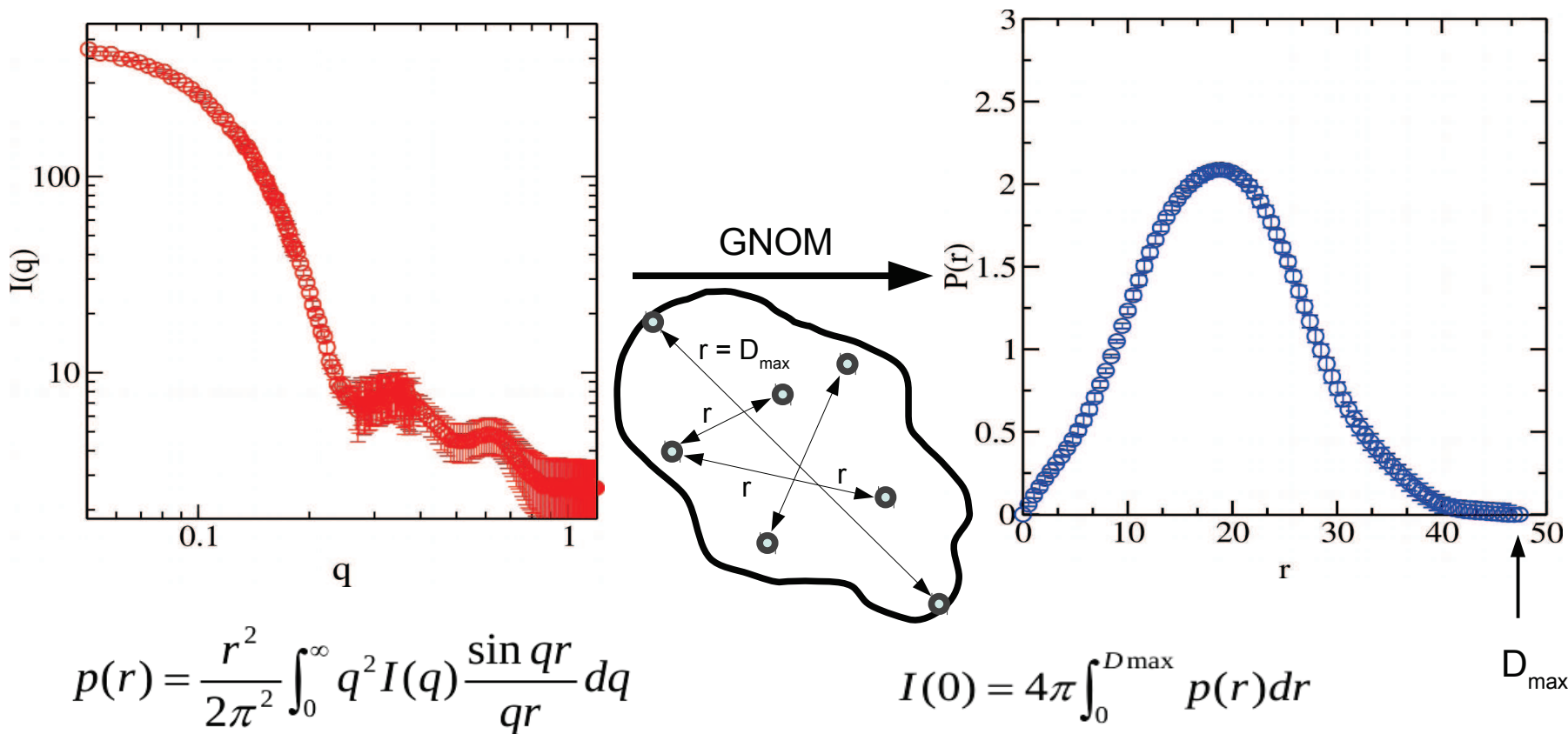
IGMCH4 Dimer



# Modeling

## Transformation of scattering curves to real space

Transformation of the scattering curve from reciprocal space to real space with the program GNOM to get the pair distance distribution function  $p(r)$  (PDDF)

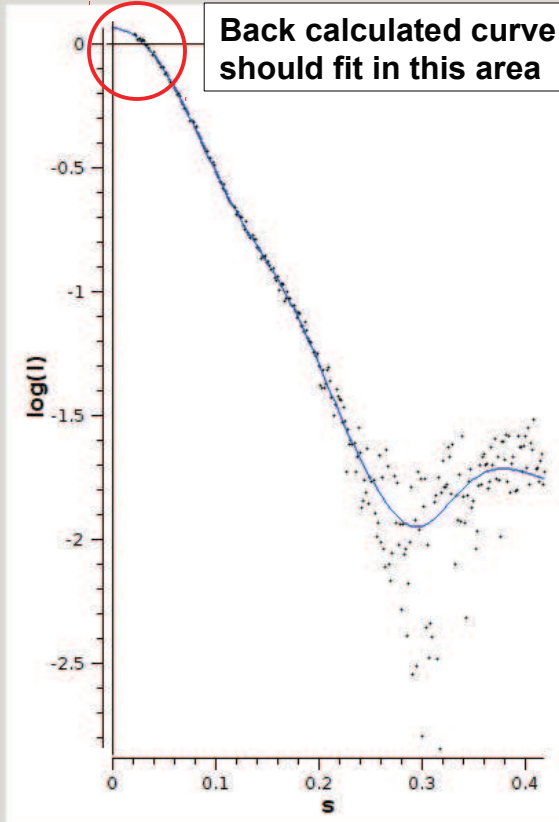


## Transformation of scattering curves to real space with GNOM

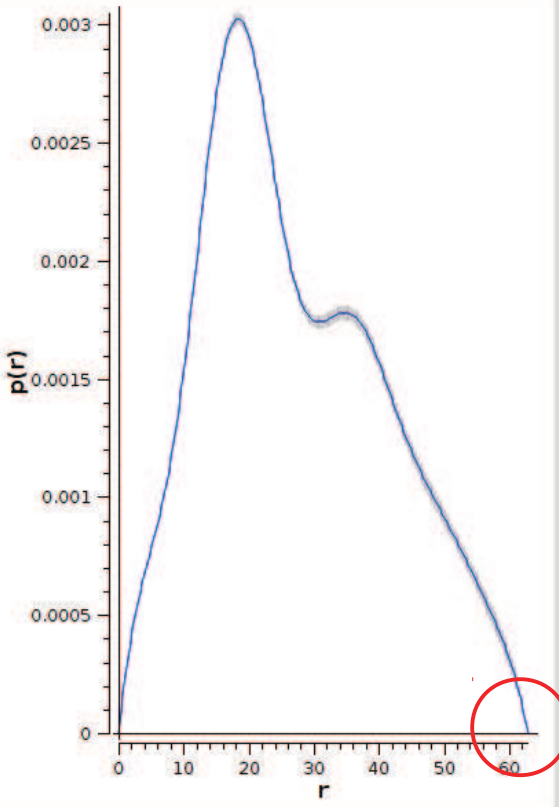
Primus Distance Distribution Wizard

Distance Distribution Analysis  
/root/saxs-treat/20131114|H-nemo\_1/export/diubiquitin\_code\_9.9mg\_subtr.dat

Experimental and back calculated  $R_g$  should be the similar



Back calculated curve should fit in this area



Quality	71.75
Experimental $R_g/l_0$	21.66 1.16
$p(r) R_g/l_0$	21.69 1.16
Porod Volume	28338.50

System: Arbitrary Monodisperse

Range: 13 to 291

$D_{max}$ : 63.00

$R_{min}=0$    $R_{max}=0$

Points: 172

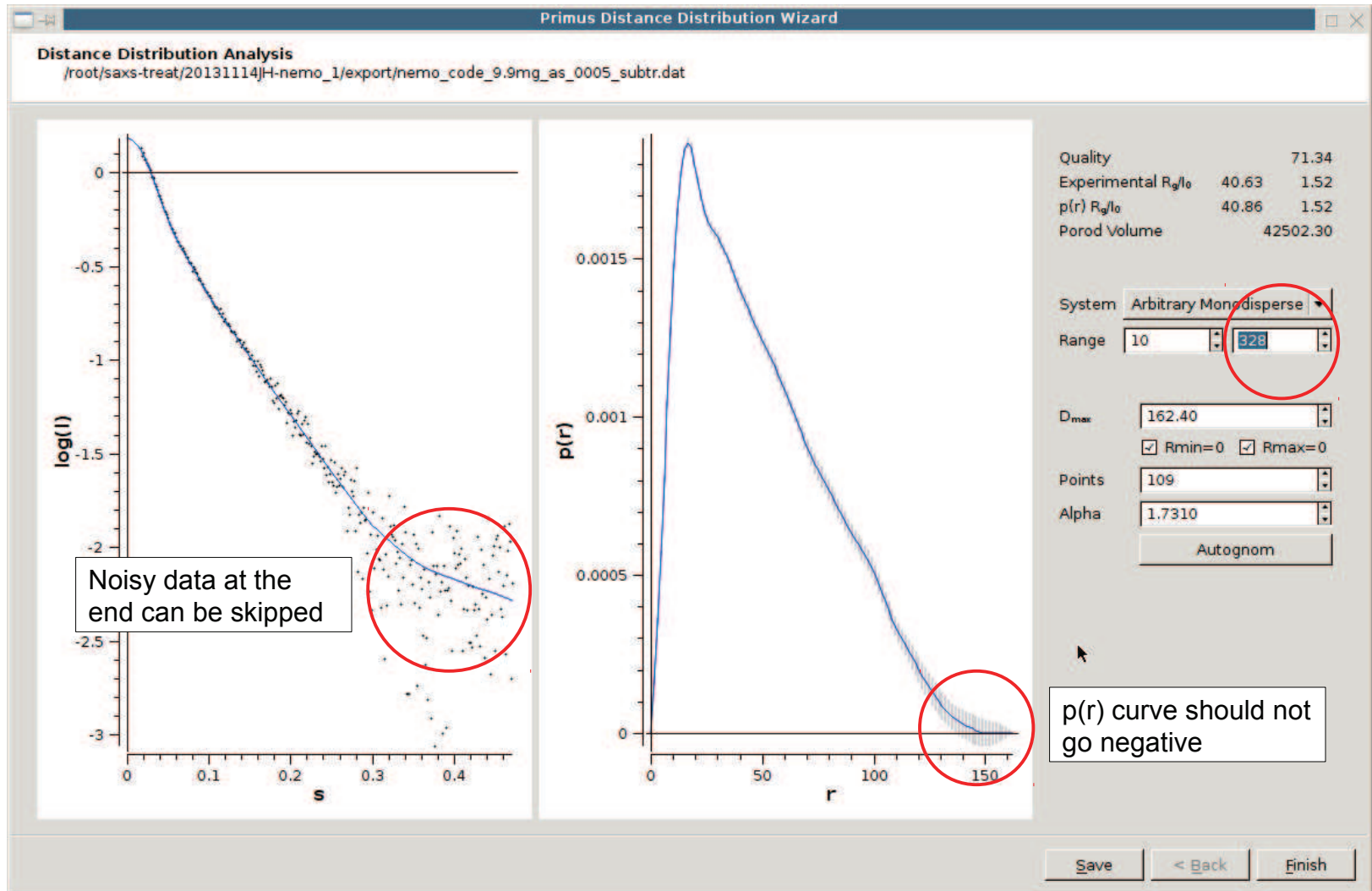
Alpha: 3.4230

Autognom

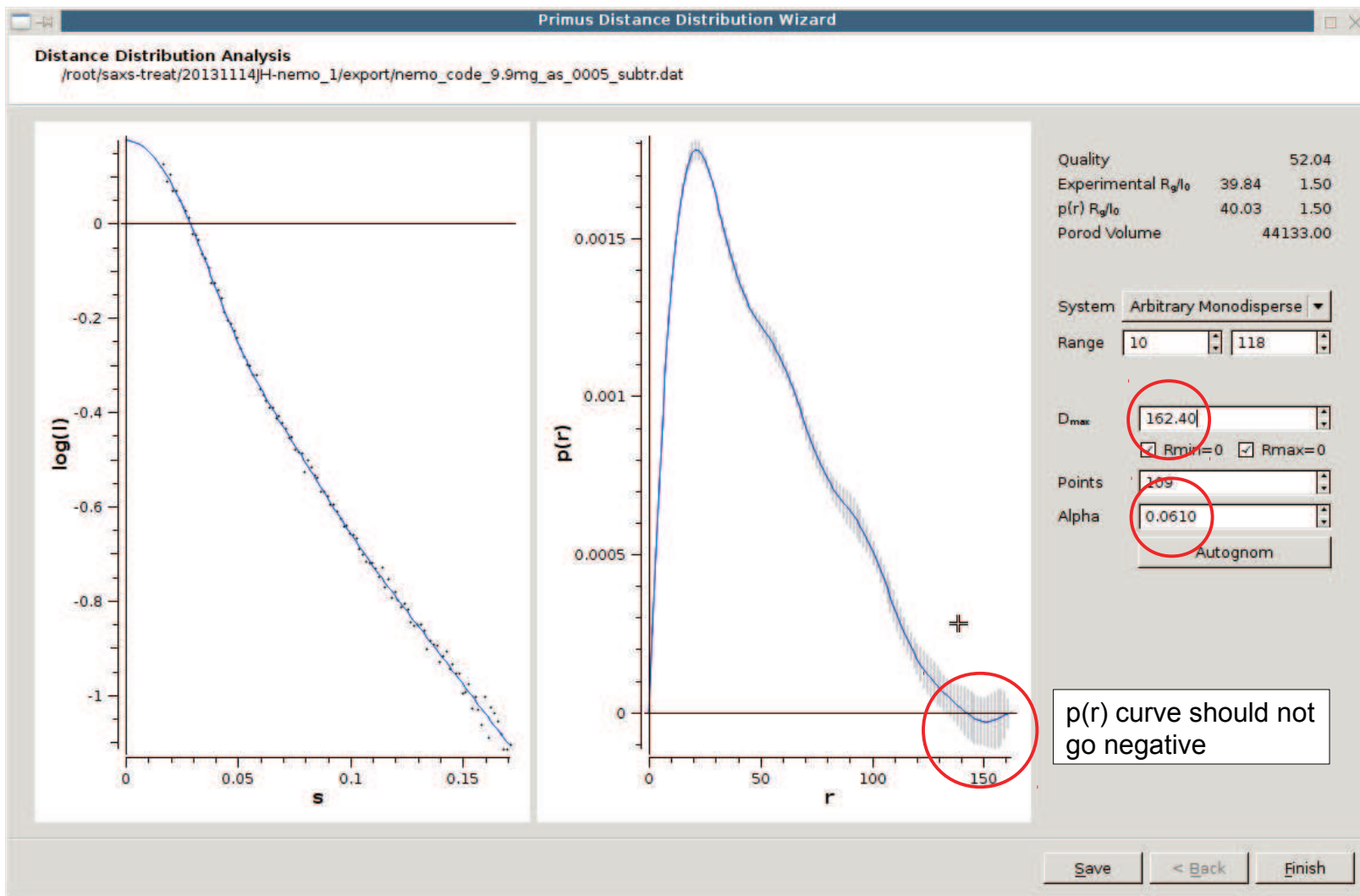
Curve should go smooth to zero (here it is too steep)

Save < Back Finish

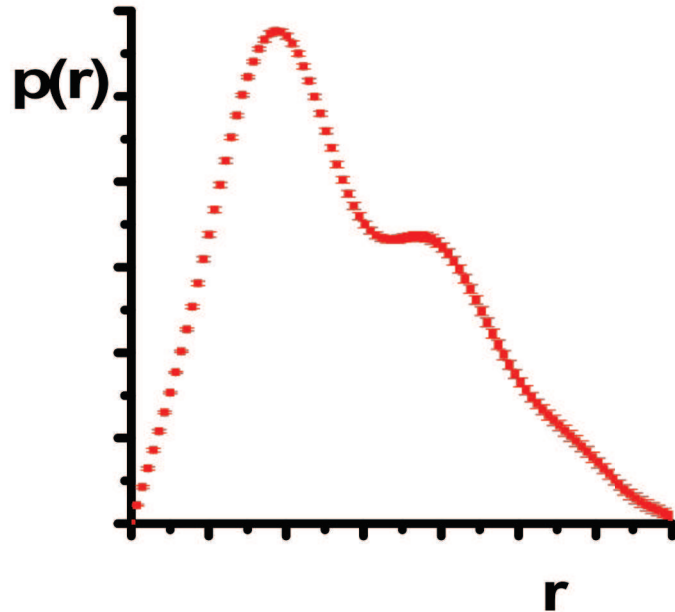
## Transformation of scattering curves to real space with GNOM



## Transformation of scattering curves to real space with GNOM



## Calculating ab initio models

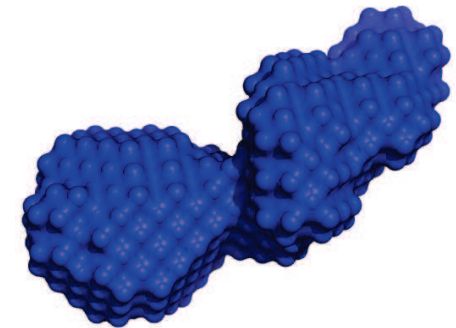
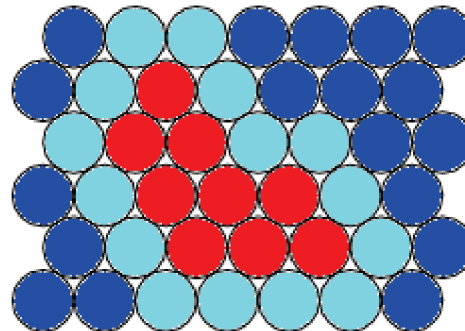
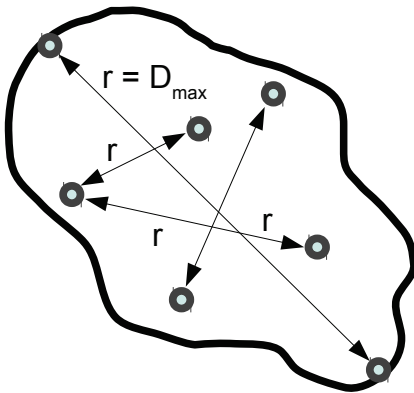


**Dammin** initially 2000 beads fixed Volume. Each bead can be flipped. Lots of symmetries allowed.

**Dammif** flexible Volume. Only beads at the phase border can flip. Few symmetries.

Both have problems with flat disks. To avoid this force symmetry (P5, P6)

10-20 models are calculated normally. They are superimposed merged and the result is filtered to the excluded volume of the particle.

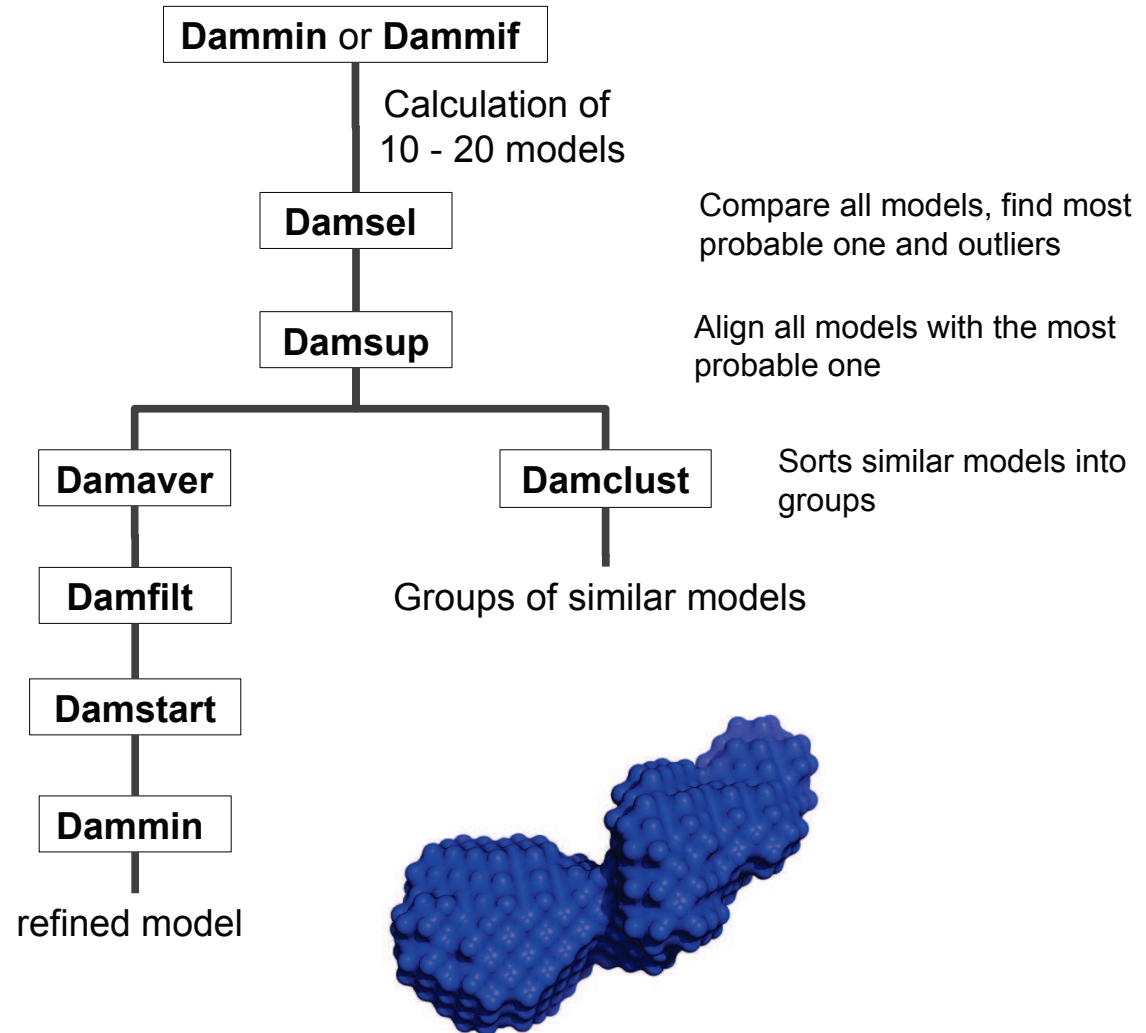


Ab initio shape determination by simulated annealing using a single phase dummy atom model

Average aligned models and compute probability map

Filters the averaged model at a given cut-off volume

Generates an input file from the averaged model with fixed core for dammin



# Rigid body modeling

## Fit of partial structures to SAXS data

**sasref** fit of complexes. Input multiple chains. Symetry and distance constrains possible.

**coral** fit of multi domain proteins. Lines up domains to one chain. Flexible linkers are taken from a database. Symetry and distance constrains possible.

**bunch** fit of multi domain proteins. Needs a sequence including missing and flexible parts. Domains are rotated around residues of flexible parts. Distance constrains possible.

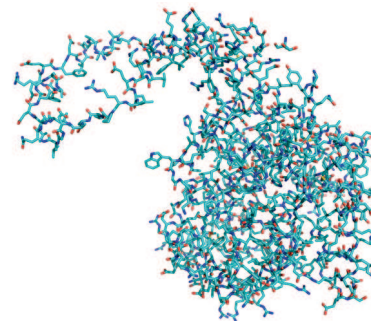
**eom** ensemble calculation. Produces 10000 structures clusters them to “chromosomes” and fits them with an evolutionary approach to the experimental data. Lines up domains to one chain. Flexible linkers have to be defined. Symetry and distance constrains possible in a limited way.

## Fit of partial structures to SAXS data: sasref

```

$ sasref
=== SASREF06 started at      21-May-2015  10:42:49
Computation mode ..... : User
Project identifier ..... : c9
Project description ..... : c9
Random sequence initialized from ..... : 104249989
Total number of subunits ..... : 1
Point symmetry of the particle ..... : P222
Number of equivalent positions ..... : 4
Total number of scattering curves ..... : 1
1-st construct, of the first and the last subunits: 1,1
File name, 1-st experimental data ..... : sub_004.dat
Number of experimental points found ..... : 1370
Angular units multiplied by ..... : 0.1000
Experimental radius of gyration ..... : 248.0
Number of points in the Guinier Plot ..... : 5
Amplitudes, 1-st subunit ..... : mon00.alm
Number of points in partial amplitudes ..... : 51
Maximum order of harmonics ..... : 15
Evaluated from the file ..... : mon.pdb
Current subunit: 3089 atoms read, center at  0.03 -0.02  0.11
Spatial step in angstroms ..... : 5.000
Angular step in degrees ..... : 20.00
Cross value ..... : 68.20
Dock value ..... : 0.0
Discontiguity value ..... : 0.0
Cross penalty weight ..... : 10.00
Disconnectivity penalty weight ..... : 10.00
Docking penalty weight ..... : 0.5000
Expected particle anisometry ..... : Unknown
.
.

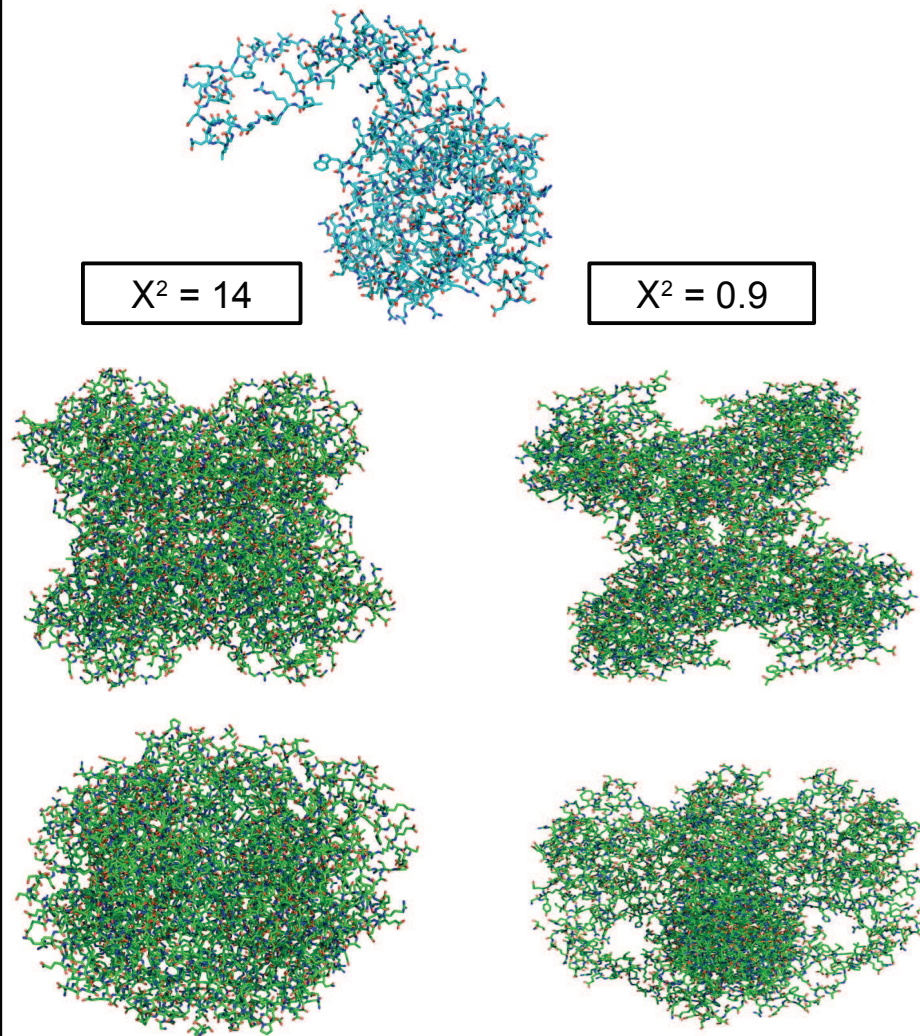
```



## Fit of partial structures to SAXS data: sasref

```

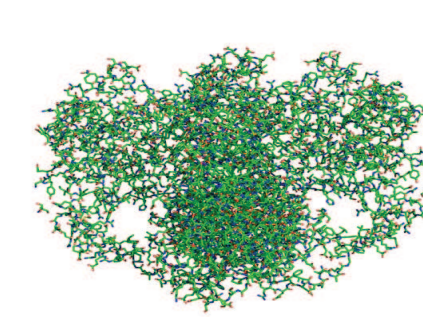
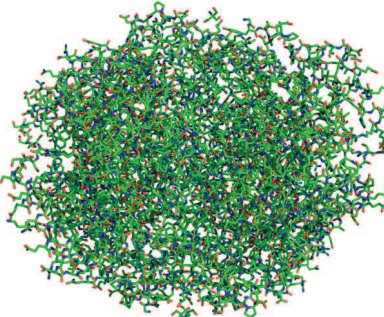
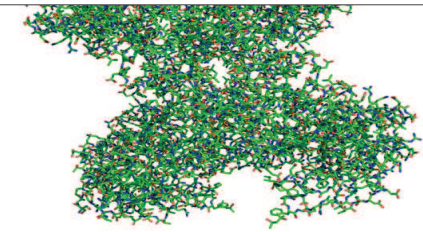
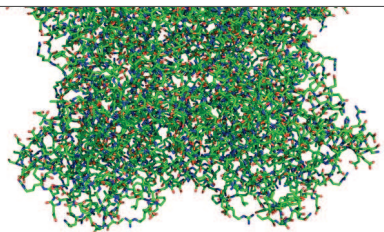
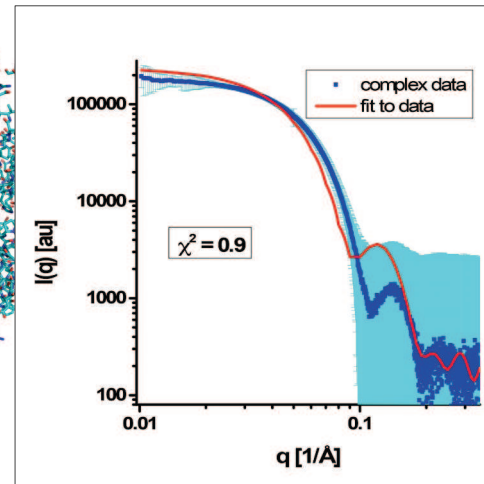
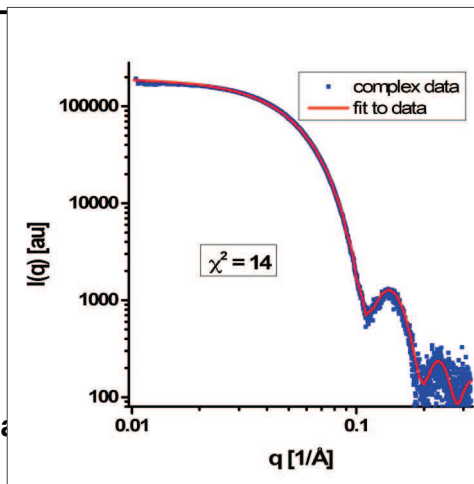
$ sasref
=== SASREF06 started at      21-May-2015  10:42:49
Computation mode ..... : User
Project identifier ..... : c9
Project description ..... : c9
Random sequence initialized from ..... : 104249989
Total number of subunits ..... : 1
Point symmetry of the particle ..... : P222
Number of equivalent positions ..... : 4
Total number of scattering curves ..... : 1
1-st construct, of the first and the last subunits: 1,1
File name, 1-st experimental data ..... : sub_004.dat
Number of experimental points found ..... : 1370
Angular units multiplied by ..... : 0.1000
Experimental radius of gyration ..... : 248.0
Number of points in the Guinier Plot ..... : 5
Amplitudes, 1-st subunit ..... : mon00.alm
Number of points in partial amplitudes ..... : 51
Maximum order of harmonics ..... : 15
Evaluated from the file ..... : mon.pdb
Current subunit: 3089 atoms read, center at  0.03 -0.02  0.11
Spatial step in angstroms ..... : 5.000
Angular step in degrees ..... : 20.00
Cross value ..... : 68.20
Dock value ..... : 0.0
Discontiguity value ..... : 0.0
Cross penalty weight ..... : 10.00
Disconnectivity penalty weight ..... : 10.00
Docking penalty weight ..... : 0.5000
Expected particle anisometry ..... : Unknown
    
```



## Fit of partial structures to SAXS data: sasref

```

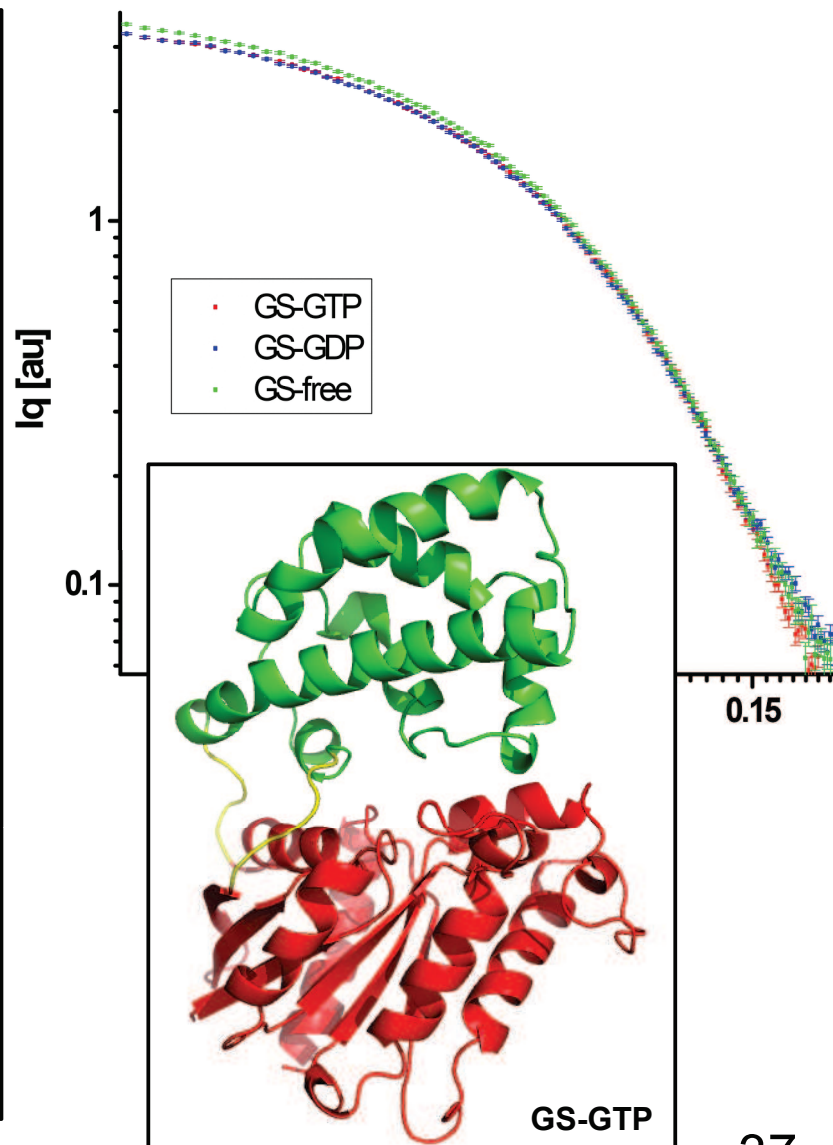
$ sasref
=== SASREF06 started at      21-May-2015  10:42:49
Computation mode ..... : User
Project identifier ..... : c9
Project description ..... : c9
Random sequence initialized from ..... : 104249989
Total number of subunits ..... : 1
Point symmetry of the particle ..... : P222
Number of equivalent positions ..... : 4
Total number of scattering curves ..... : 1
1-st construct, of the first and the last subunits: 1,1
File name, 1-st experimental data ..... : sub_004.d
Number of experimental points found ..... : 1370
Angular units multiplied by ..... : 0.1000
Experimental radius of gyration ..... : 248.0
Number of points in the Guinier Plot ..... : 5
Amplitudes, 1-st subunit ..... : mon00.alm
Number of points in partial amplitudes ..... : 51
Maximum order of harmonics ..... : 15
Evaluated from the file ..... : mon.pdb
Current subunit: 3089 atoms read, center at  0.03 -0.02  0.11
Spatial step in angstroms ..... : 5.000
Angular step in degrees ..... : 20.00
Cross value ..... : 68.20
Dock value ..... : 0.0
Discontiguity value ..... : 0.0
Cross penalty weight ..... : 10.00
Disconnectivity penalty weight ..... : 10.00
Docking penalty weight ..... : 0.5000
Expected particle anisometry ..... : Unknown
    
```



## Fit of partial structures to SAXS data: coral

```

$ coral
=== CORAL05 started at      23-Apr-2015  19:35:35
Computation mode ..... : User
Project identifier ..... : 2-0a
Project description ..... : 2-0a
Random sequence initialized from ..... : 193535
File name with objects info ..... : c.con
Coordinates of the 1-st subunit evaluated from ..... : RED1.pdb
 198 atoms read, center at  26.89 -26.80  8.09
Subunit was fixed
Coordinates of the 2-nd subunit evaluated from ..... : GREEN.pdb
 911 atoms read, center at  52.99 -19.12 12.51
Coordinates of the 3-rd subunit evaluated from ..... : RED2.pdb
1386 atoms read, center at  31.86 -35.37  1.03
Subunit was fixed
Number of backbone atoms generated ..... : 316
Averaged formfactors of DRs used
DR formfactor multiplier ..... : 1.000
Point symmetry of the particle ..... : P1
Number of equivalent positions ..... : 1
Cross penalty ..... : 0.3349
Cross penalty weight ..... : 100.0
Shift penalty ..... : 2.376
Shift penalty weight ..... : 1.000
Total number of scattering curves ..... : 1
1-st construct, the first and the last residues: 1 and 316
File name, 1-st experimental data ..... : GS-GTP-10mg.dat
Number of experimental points found ..... : 481
Experimental radius of gyration ..... : 22.20
    
```

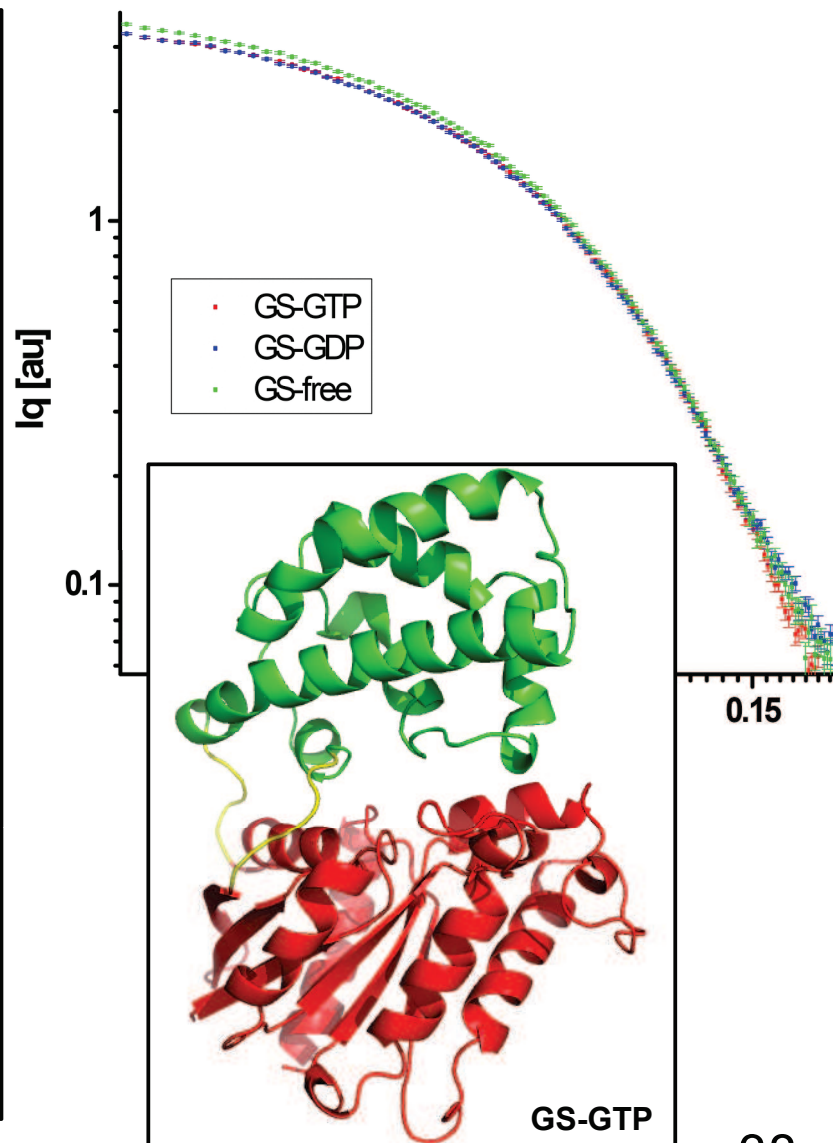


## Fit of partial structures to SAXS data: coral

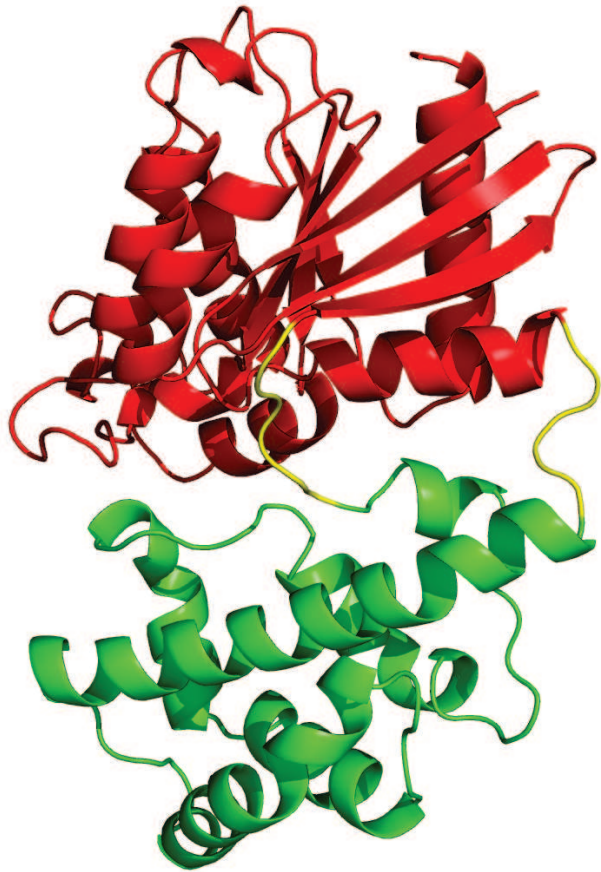
```
$ coral
=== CORAL05 started at 23-Apr-2015 19:35:35
Computation mode ..... : User
Project identifier ..... : 2-0a
Project description ..... : 2-0a
Random sequence initialized from ..... : 193535
File name with objects info ..... : c.con
Coordinates of the 1-st subunit evaluated from ..... : RED1.pdb
198 atoms read, center at 26.89 -26.80 8.09
Subunit was fixed
Coordinates of the 2-nd subunit evaluated from ..... : GREEN.pdb
011 atoms read, center at 52.00 10.12 12.51
```



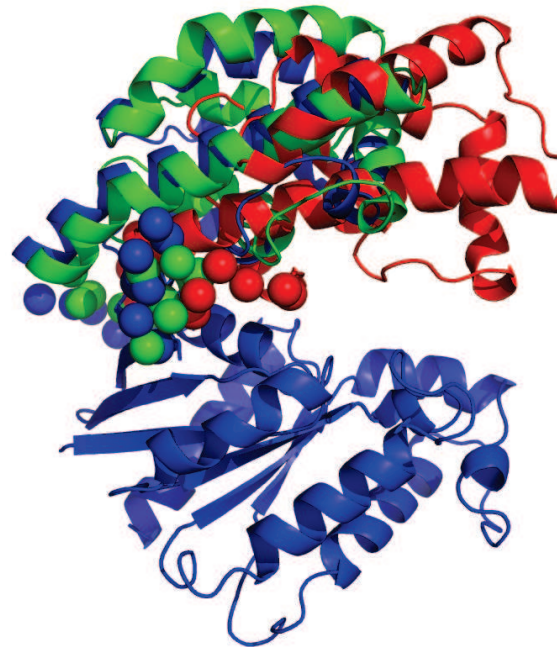
GS-free



## Fit of partial structures to SAXS data: coral

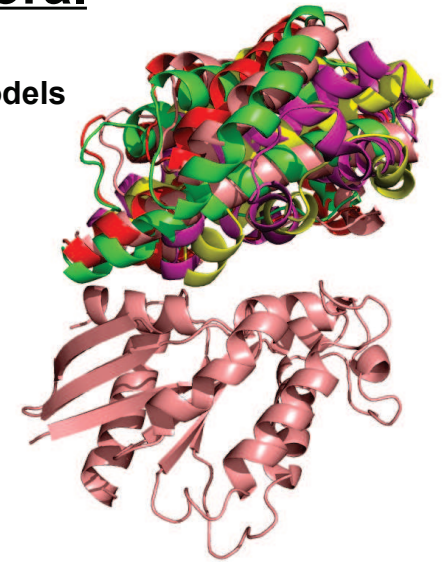


GS-GTP crystal



GS-GTP red  
GS-GDP green  
GS-free blue

GS-GTP 5 models

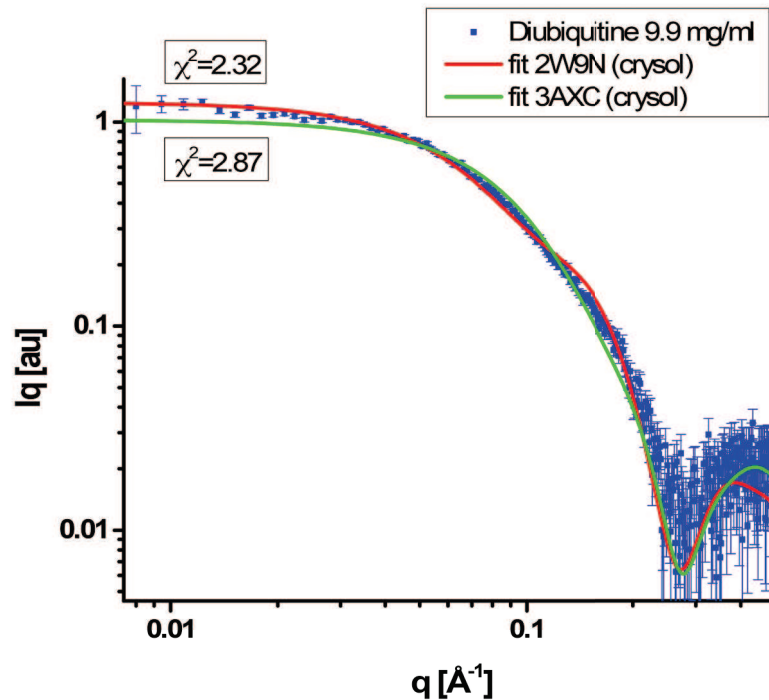


GS-GTP 5 models red  
GS-GDP 5 models green

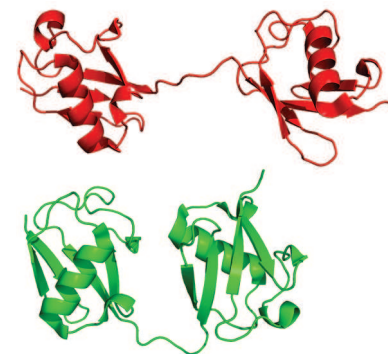
## Fit of pieces of structures to SAXS data: eom

```

$ eom
  RANCH Version 2.0
  Started: ..... : Fri Oct 17 16:06:44 2014
  iSeed ..... : 1934479814
  Chain type ..... : Random
  Sequence file name: ..... : sequence.seq
  Symmetry: ..... : P1
  Symmetry type: ..... : aSymmetric
  Number of residues: ..... : 1142
  Number of domains: ..... : 2
  -----
  Domain number: ..... : 1
  Path: ..... : diubi1.pdb
  Fixed: ..... : 0
  Oligomer: ..... : 0
  DNA file name: ..... : None
  -----
  Domain number: ..... : 2
  Path: ..... : diubi2.pdb
  Fixed: ..... : 0
  Oligomer: ..... : 0
  DNA file name: ..... : None
  Contact file name: ..... : None
  Number of structures: ..... : 10000
  Suffix of the pdb files: ..... : eom
  Output Folder Path: ..... : ./
  Number of experimental curves: ..... : 1
  Experimental data filename (curve 1): ..... : diubi_9p9_subtr.dat
  .
  
```

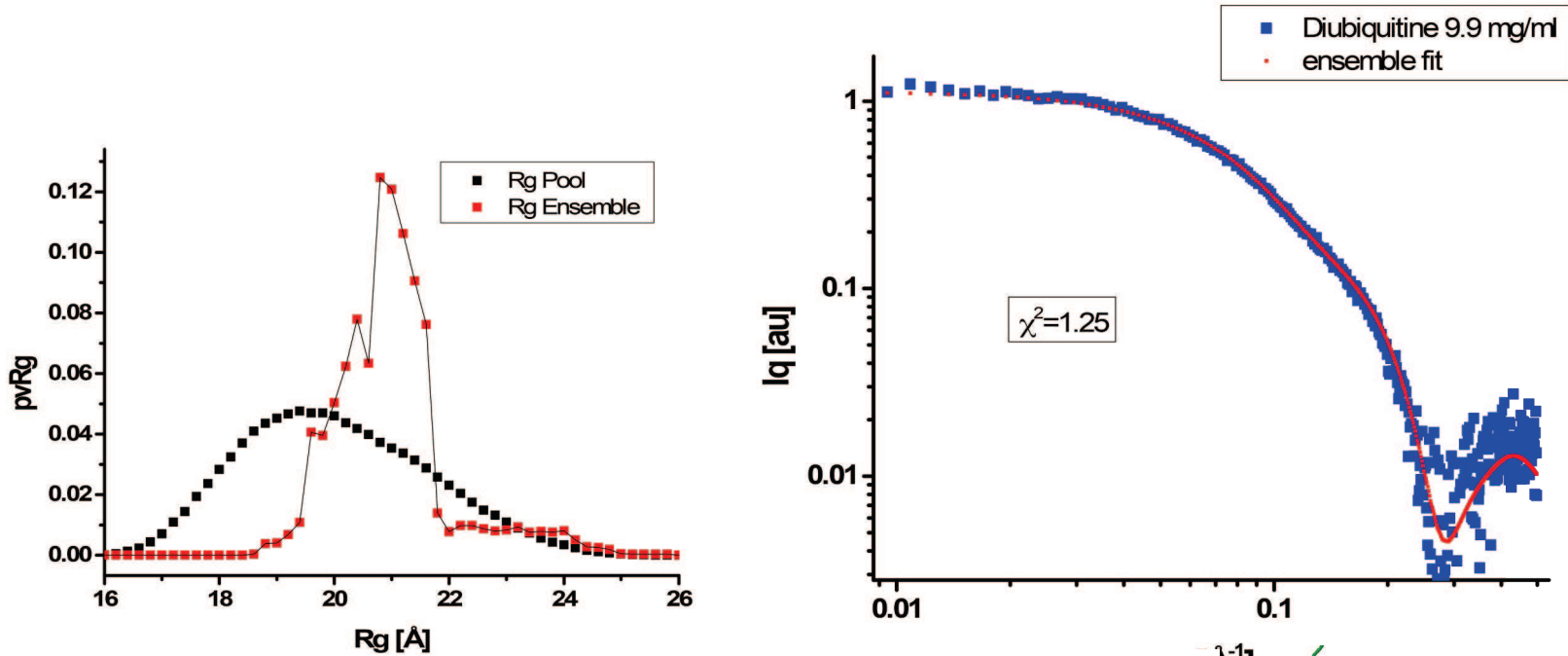


Crystal structures  
2W9N (red)  
3AXC (green)

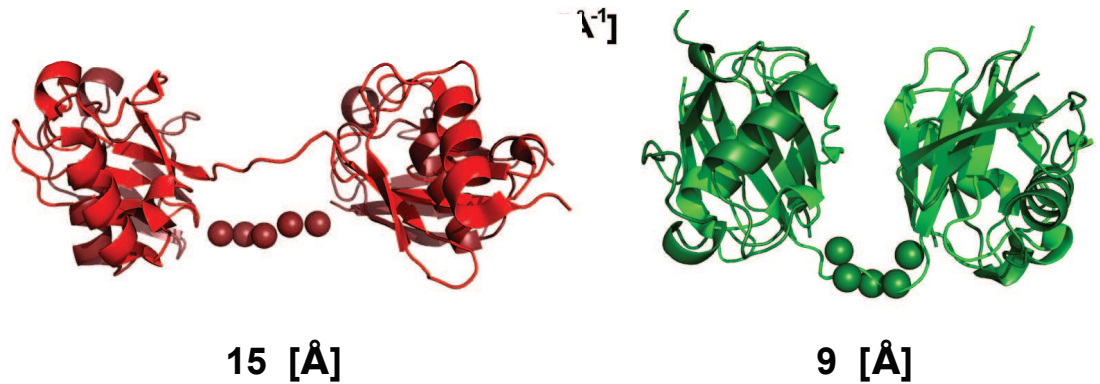


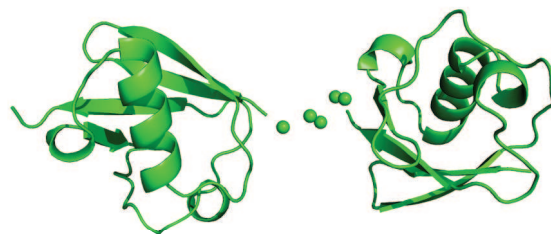
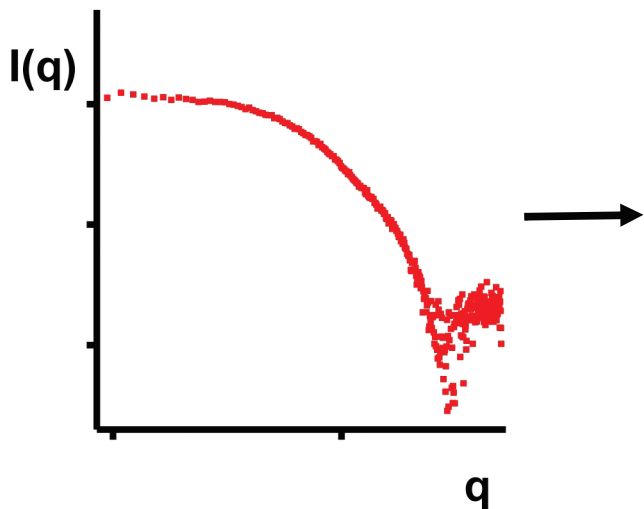
Sequence file has fasta format (without comments and chain information)

## Fit of pieces of structures to SAXS data: eom

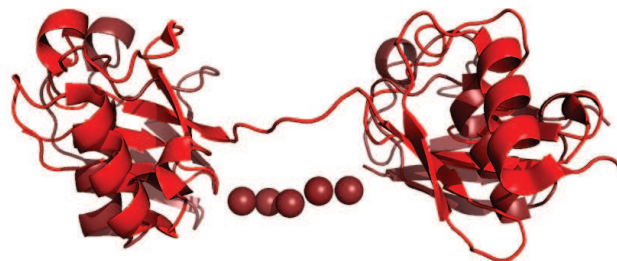


Overlay of Ensemble Models and Crystal Structures



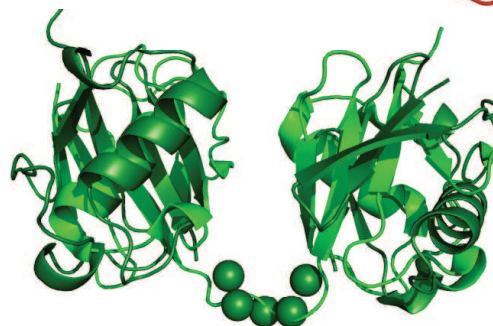


**Rigid body model**  
(average structure)



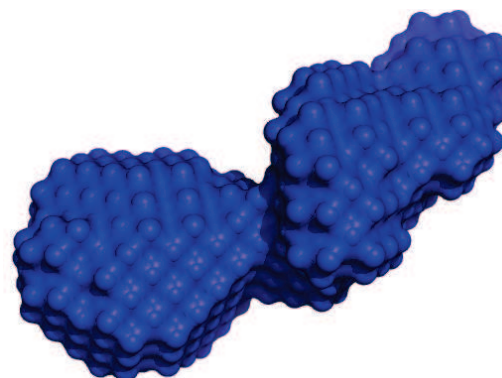
**Ensemble  
Ratio  
38:62**

(possible conformations  
of a flexible molecule)



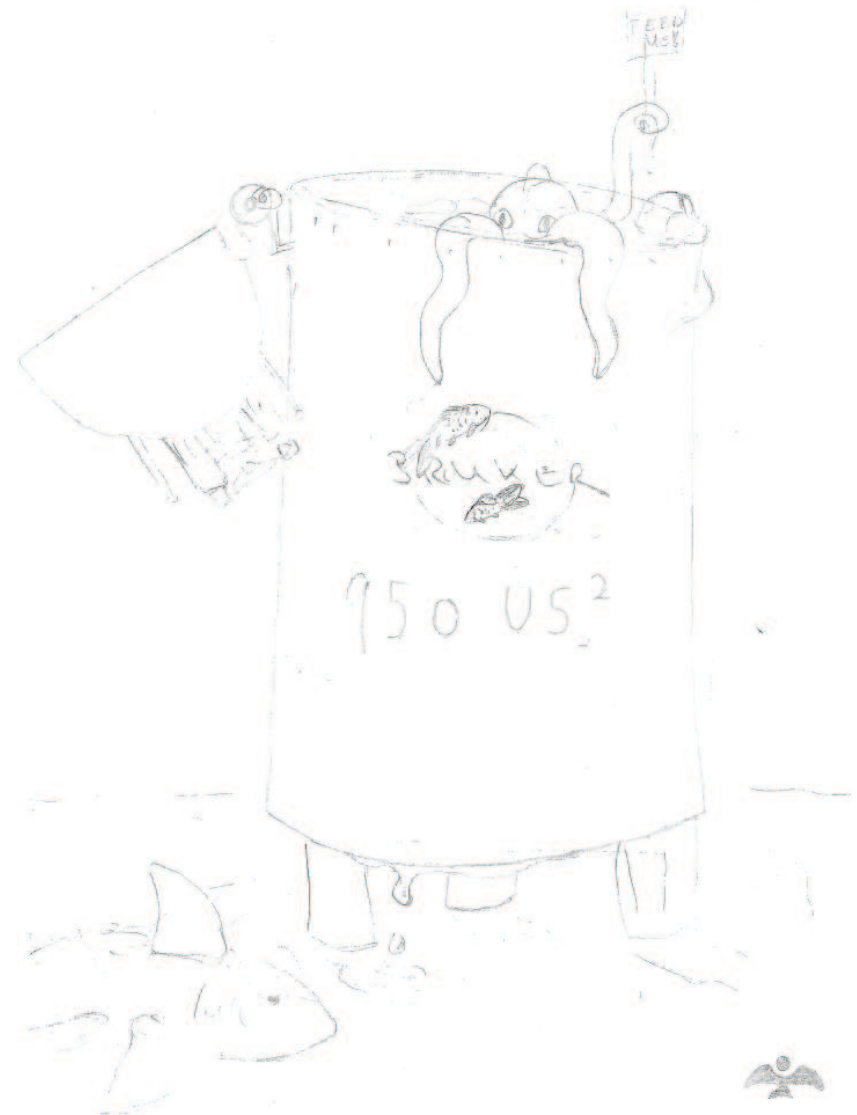
Depending on the initial assumption or question modeling of data with different software leads to different results.

All these results are correct within the limiting factors of the software

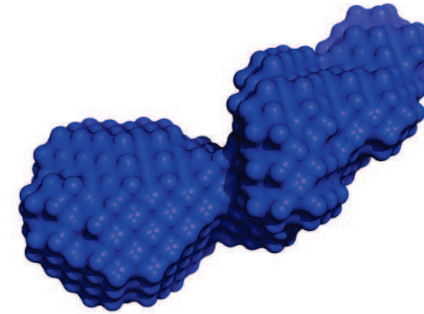
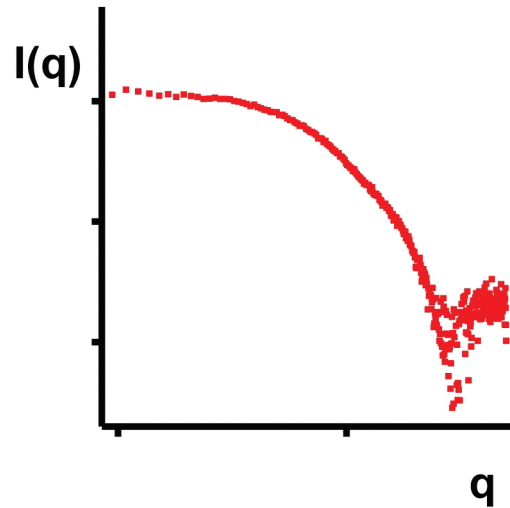


**Dummy model**  
(bead model without  
any structural  
information)

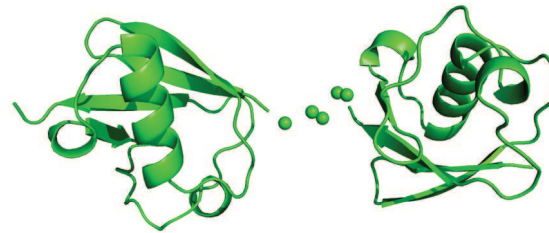
Thanks



## Rigid body modeling



Dummy model



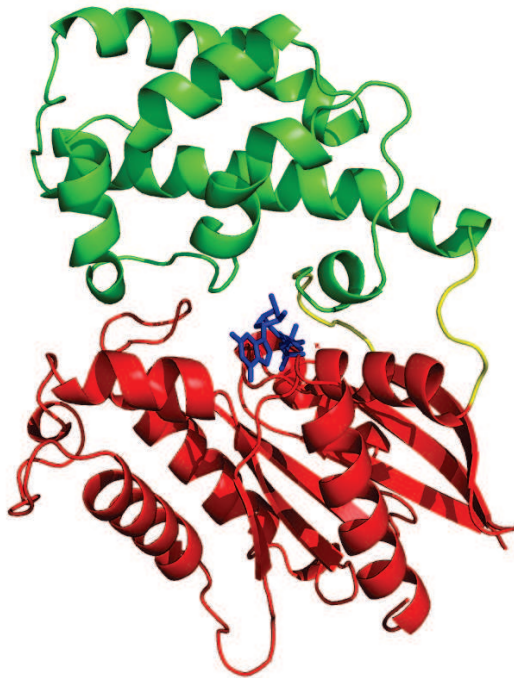
Rigid body model



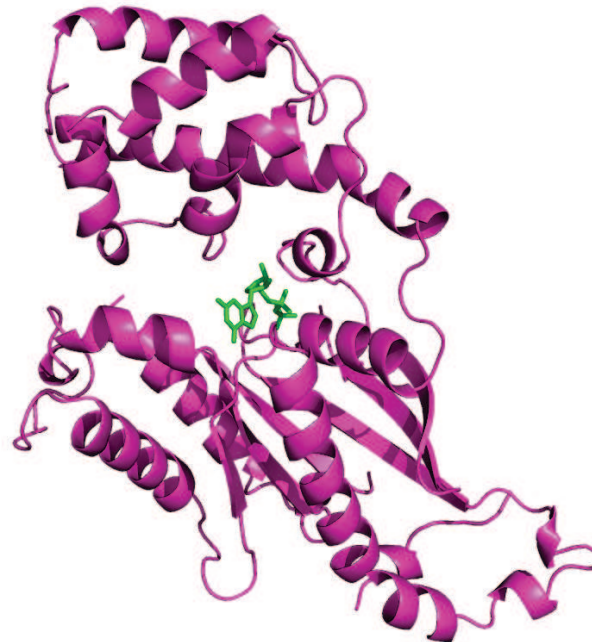
Crystal Structure

- Arrangement of domains in space

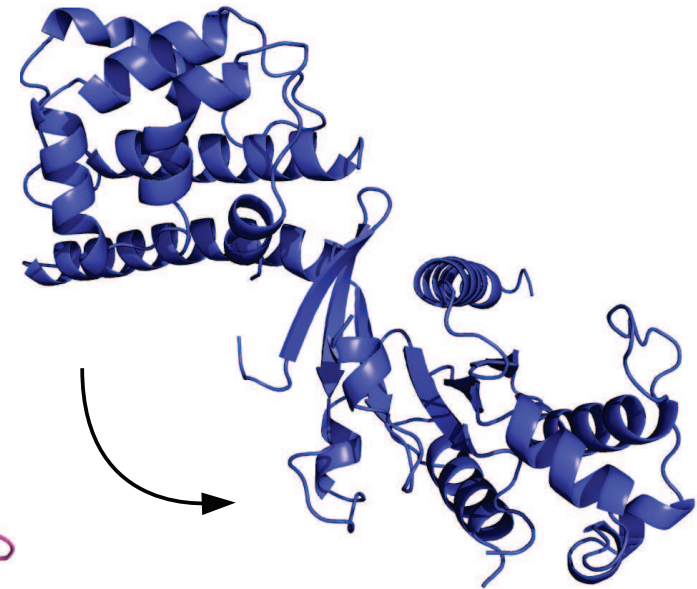
Crystal structures available for all three states



**GTP**

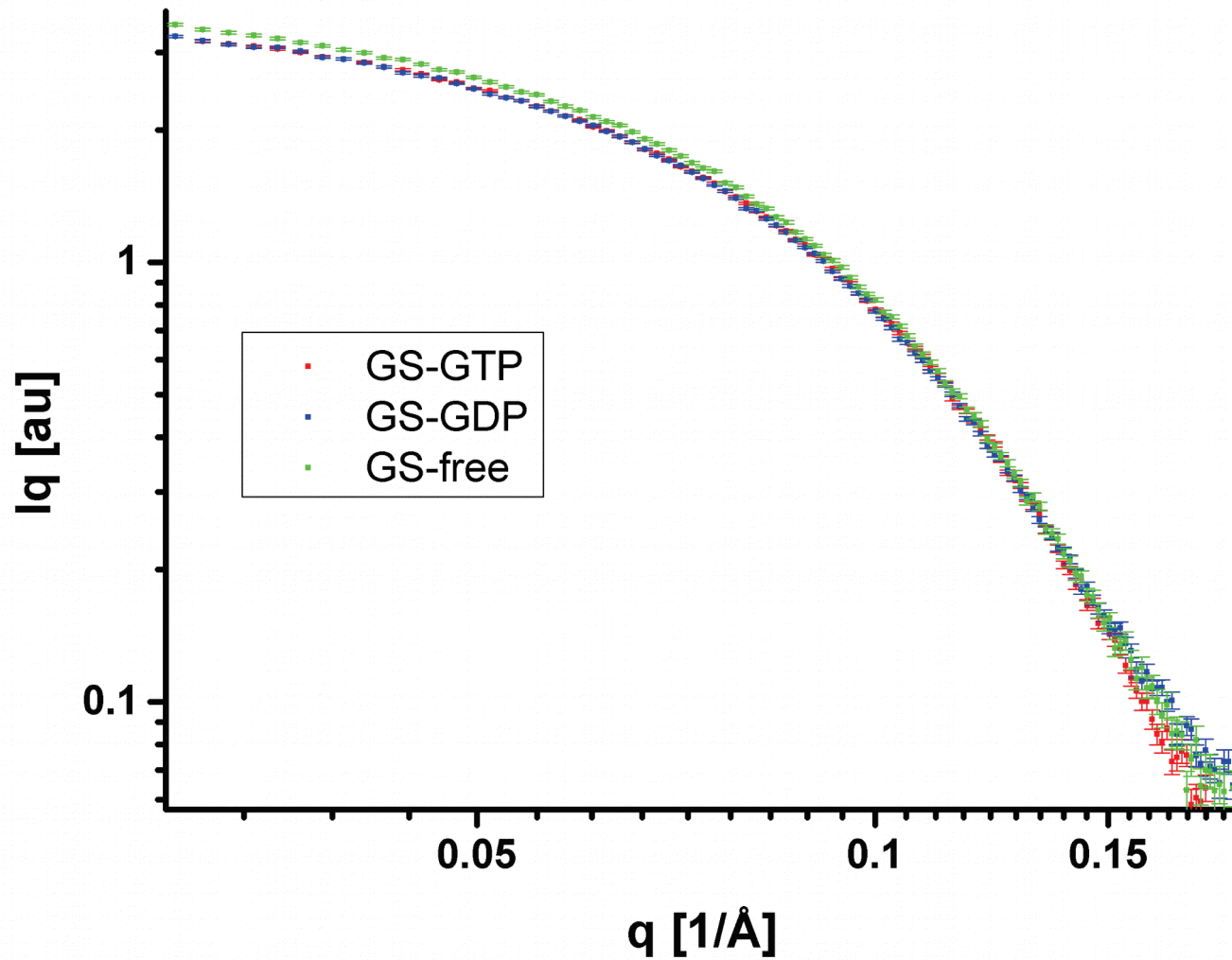


**GDP**

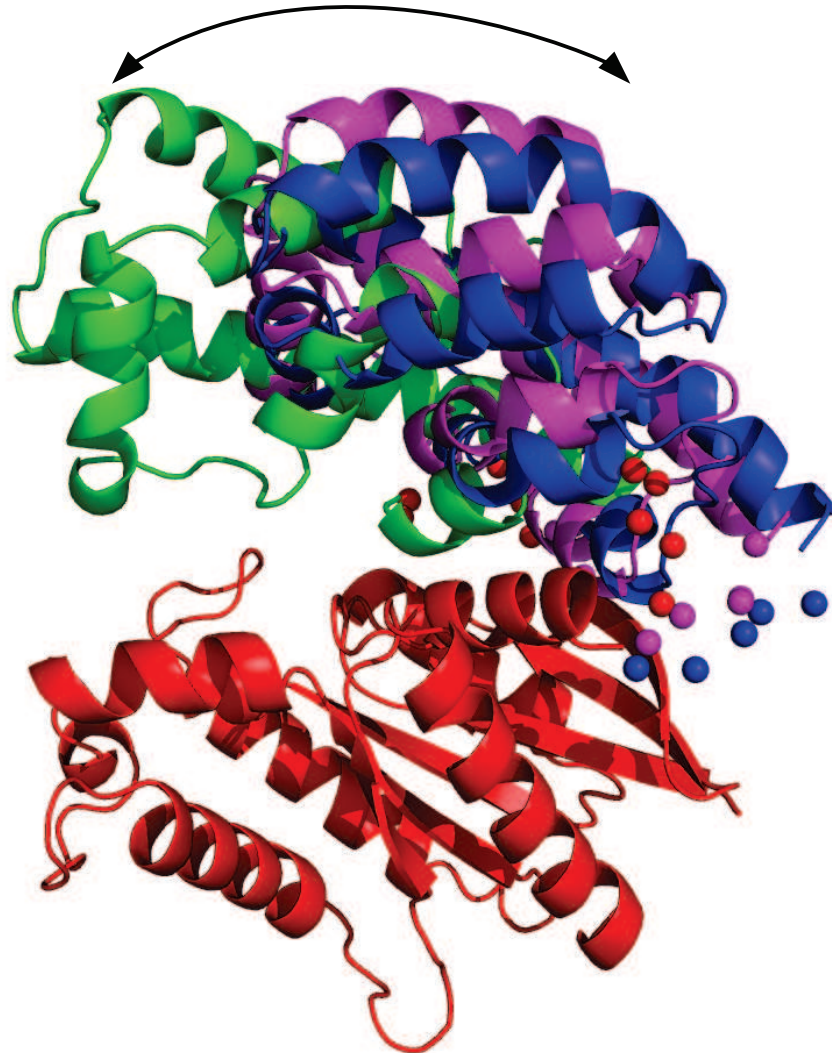


**free**

## Scattering data

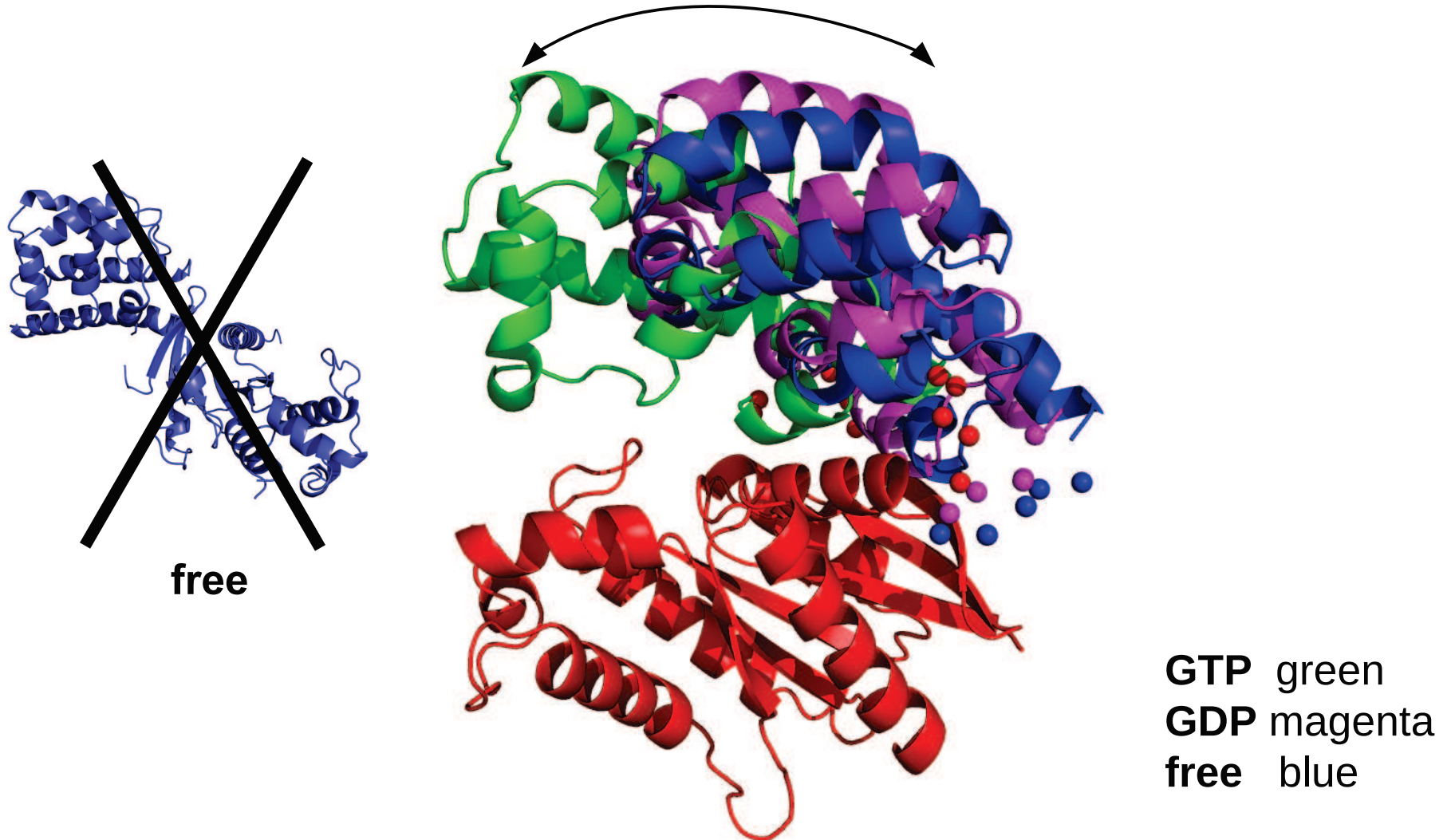


## Conformations of the G-protein in solution

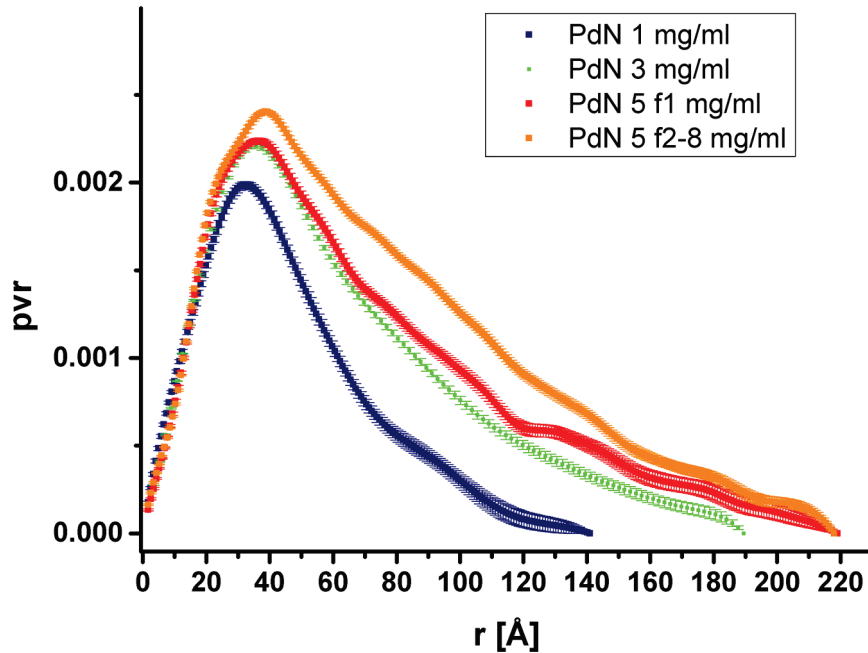


**GTP** green  
**GDP** magenta  
**free** blue

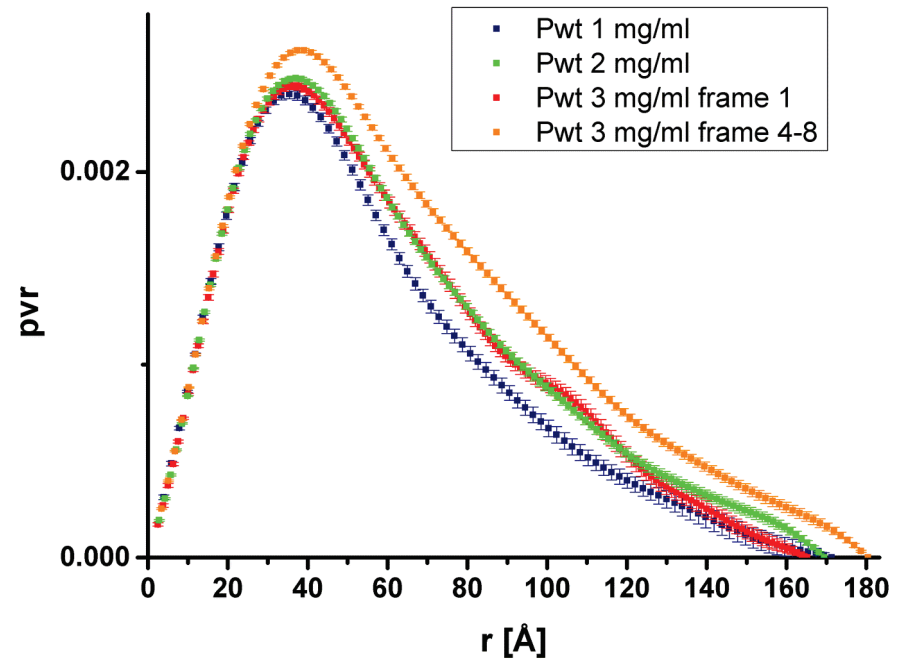
## Conformations of the G-protein in solution



Is the N-terminus unstructured?  
Is the N-terminus responsible for dimerisation?

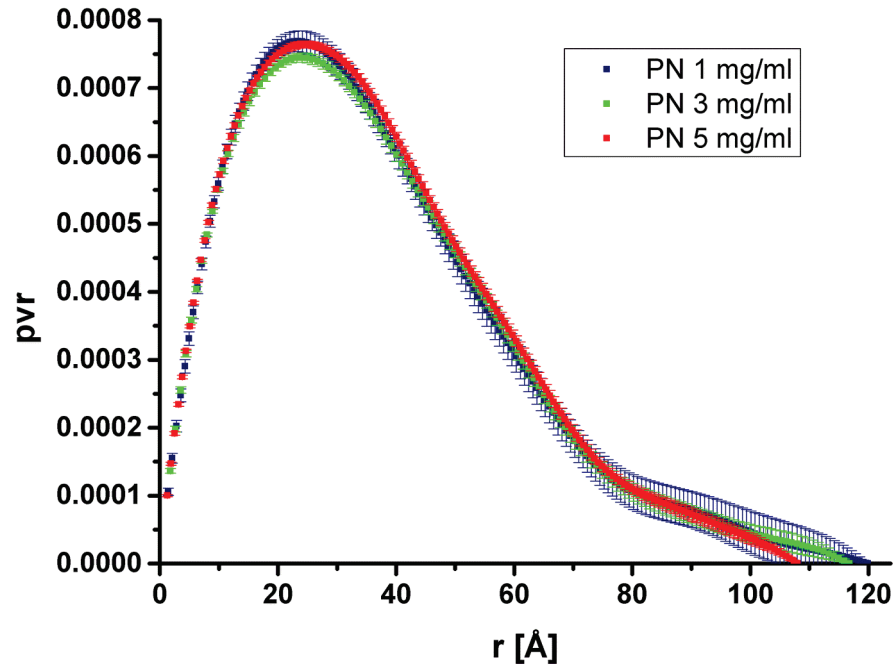


- $\delta$ N-construct seems unstable
- Concentration dependent growth of size
- Dimerisation or agglomeration



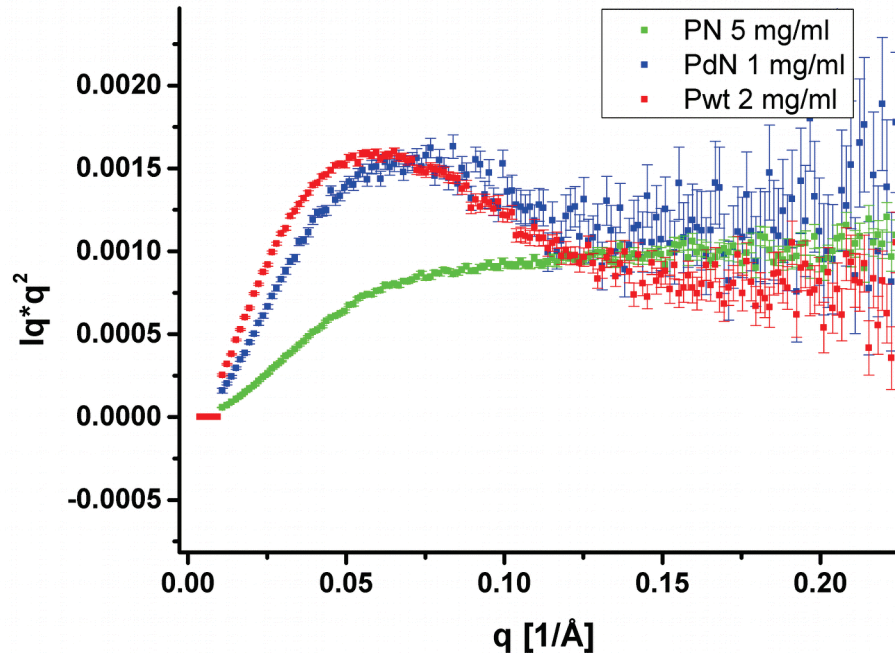
- WT is a bit more stable
- Concentration dependent growth of size
- Dimerisation or agglomeration

Is the N-terminus unstructured?  
Is the N-terminus responsible for dimerisation?



- N-terminal part stable
- No concentration dependent behaviour

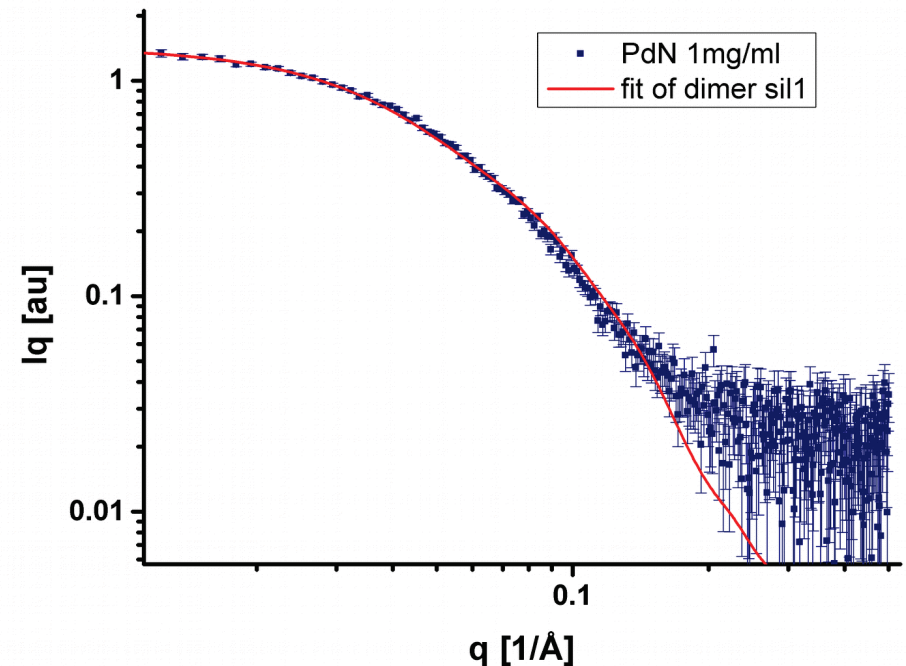
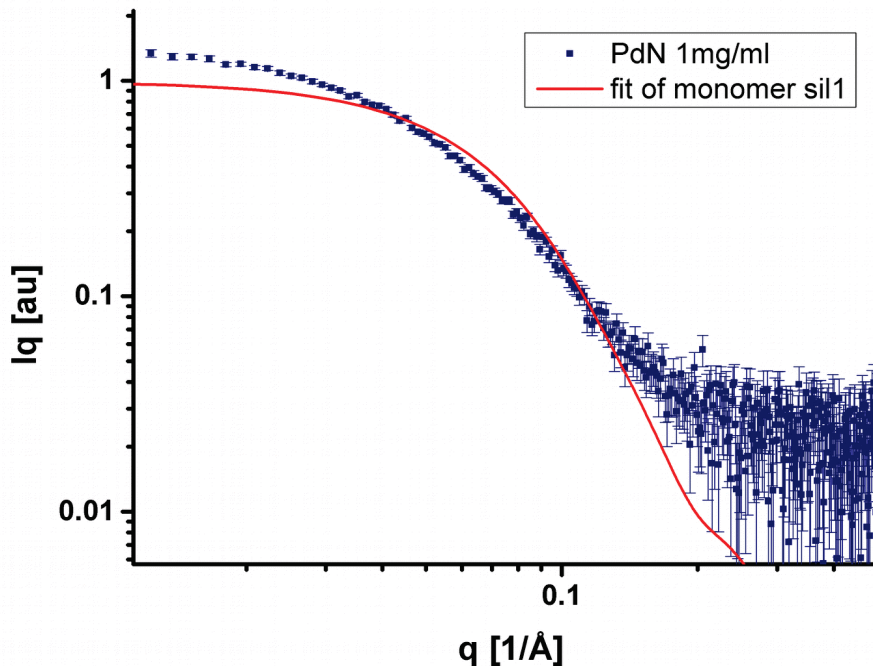
Is the N-terminus unstructured?  
Is the N-terminus responsible for dimerisation?



- WT looks more structured than  $\delta N$
- curves are too noisy artificial increase of intensity due to the noise possible

—————> The N-terminal part is unstructured

## Is the N-terminus responsible for dimerisation?



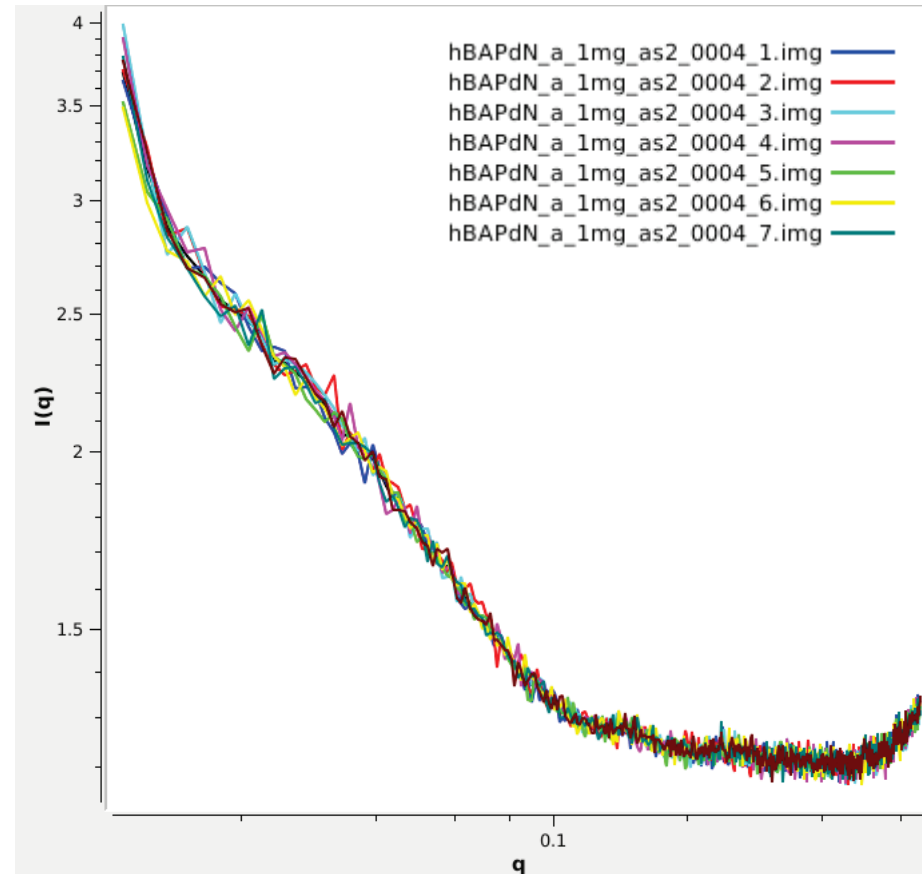
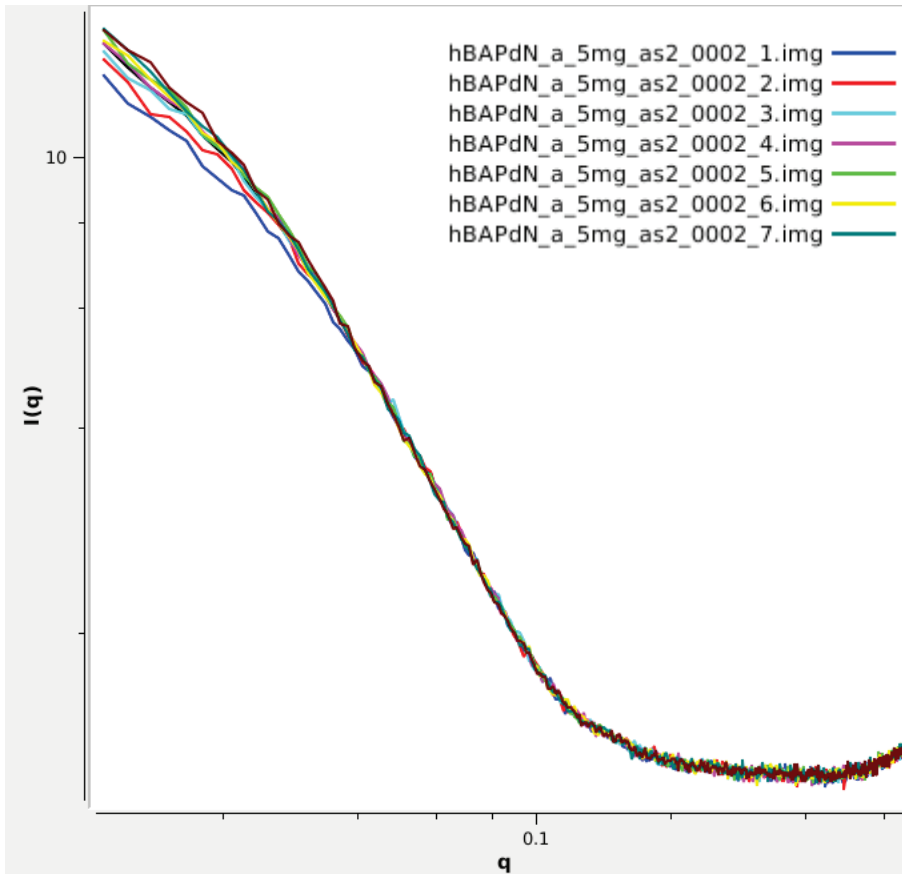
- Homology model doesn't fit at all
- Protein is bigger (higher  $I_0$ )
- Monomer is more compact (slope)

- Dimer of the Homology model fits
- $I_0$  fits
- Slope fits almost

—————> **The protein is a dimer even at the lowest concentration**

## Scattering

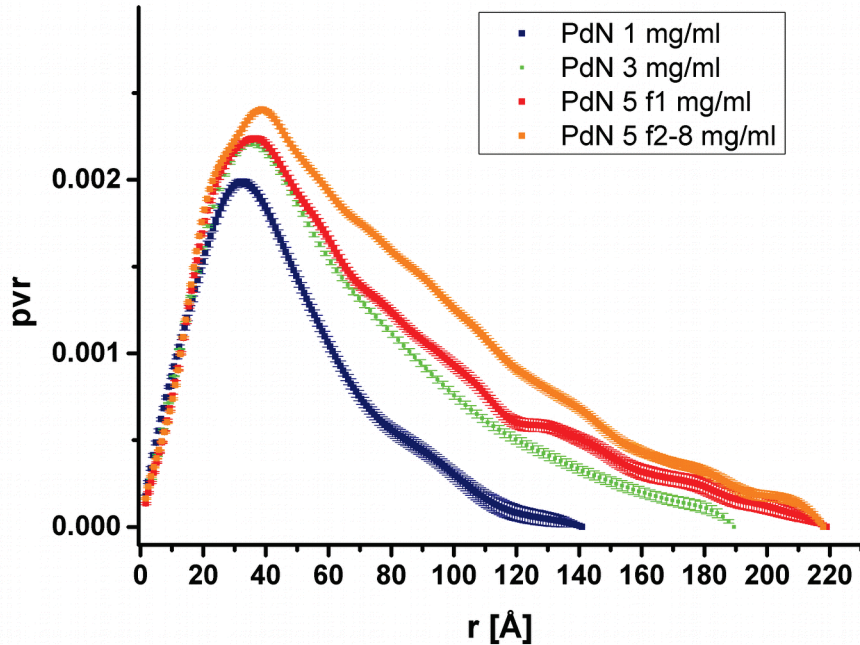
hBAPdN all frames 5mg/ml (left) 1mg/ml (right)



High concentration is affected by beamdamage.  
Low concentration has no beamdamage.

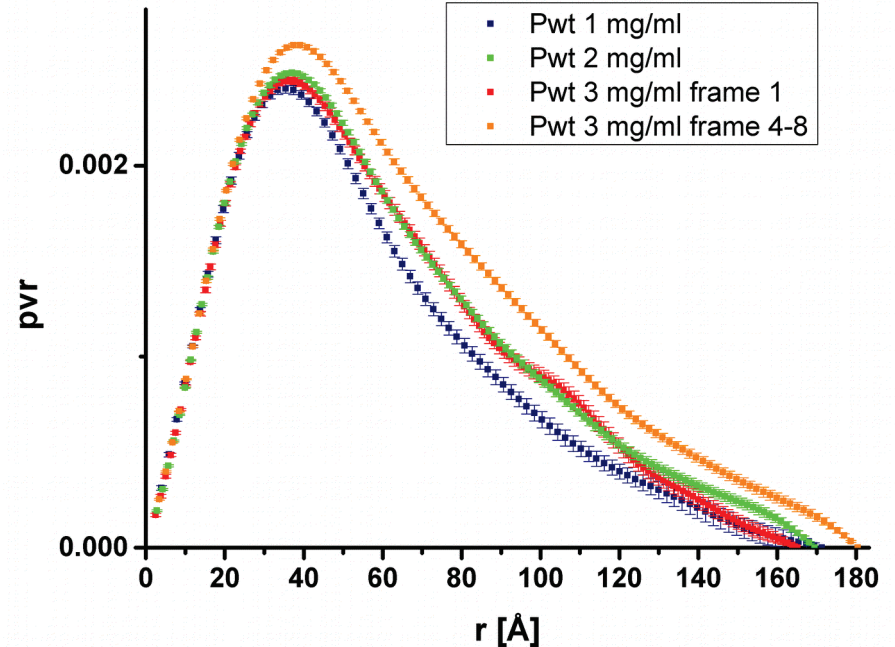
## PVR

hBAPdN



Higher concentrations have a much higher molecular weight, show beam damage and have an elongated structure. The lowest concentration has a  $D_{max}$  of  $\sim 140 \text{ \AA}$ .

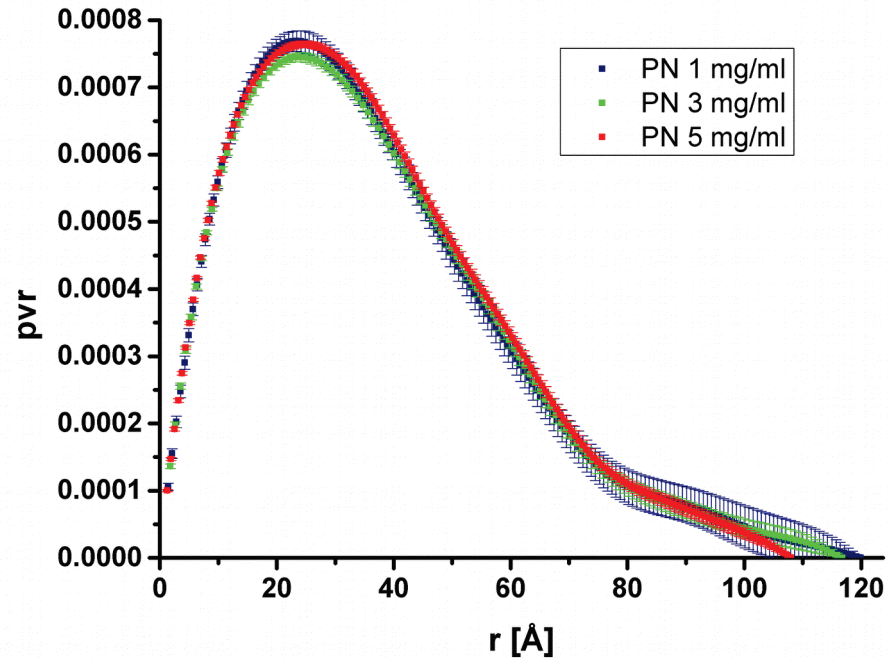
hBAPwt



The increase of molecular weight is moderate. The Structure of the Protein is elongated. All concentrations have a  $D_{max}$  around  $\sim 170 \text{ \AA}$ .

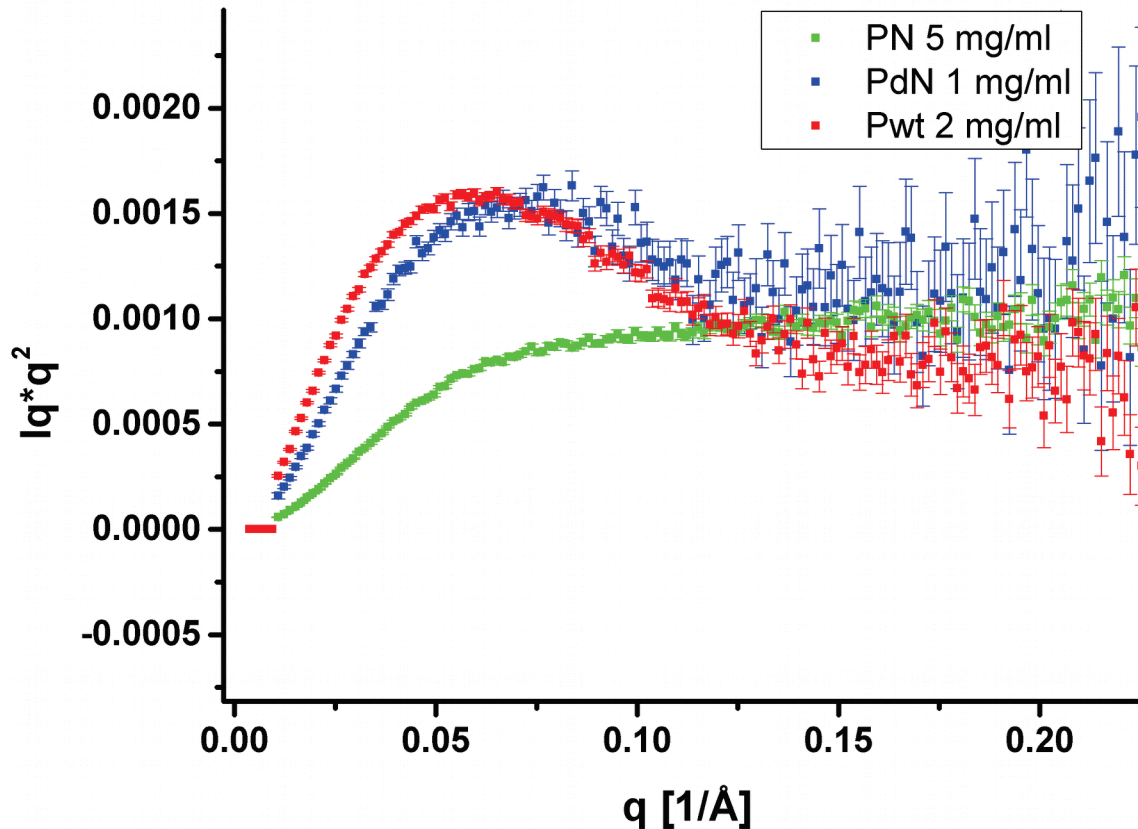
## PVR

hBAPN



The N-terminus shows no concentration dependent behaviour and has no beamdamage. All concentrations have a  $D_{max}$  around  $\sim 110$  Å.

## Kratky plot



The N-terminus is unfolded.  
 C-terminus and wt are folded.  
 The data are too noisy to see if the N-terminus in wt is structured.

## Numbers

Radius of gyration

File	Rg	+ -
hBAPdN_a_1mg_as2_cnorm.dat	36.59	1.3
hBAPdN_a_3mg_as2_cnorm.dat	49.76	0.9
hBAPdN_a_5mg_as2_0002_f4-8.dat	58.05	0.5
hBAPdN_a_5mg_as2_0002_frame1.dat	54.07	1.2
hBAPwt_b_1mg_as2_cnorm.dat	43	1.3
hBAPwt_b_2mg_as2_cnormf28.dat	47	0.8
hBAPwt_b_3mg_as2_cnorm-f1.dat	46	1.6
hBAPwt_b_3mg_as2_cnorm-f48.dat	51.9	1.0
hBAPN_c_1mg_as2_cnorm.dat	28.4	1.9
hBAPN_c_3mg_as2_cnorm.dat	29.39	0.7
hBAPN_c_5mg_as2_cnorm.dat	29.49	0.5

## Numbers

Dmax and MW

Sil1 Dmax 70 Å

File	Dmax	Vp	MW
hBAPdN_a_1mg_as2_cnorm_a	97.0	95776.0	77.6
hBAPdN_a_1mg_as2_cnorm_h	141.0	101640.0	82.3
hBAPdN_a_3mg_as2_cnorm_a	165.5	148266.0	120.1
hBAPdN_a_3mg_as2_cnorm_h	189.5	155546.0	126.0
hBAPdN_a_5mg_as2_0002_f4-8_h	218.0	248684.0	201.4
hBAPdN_a_5mg_as2_0002_frame1_h	219.0	196074.0	158.8

hBAPdN 40 kD

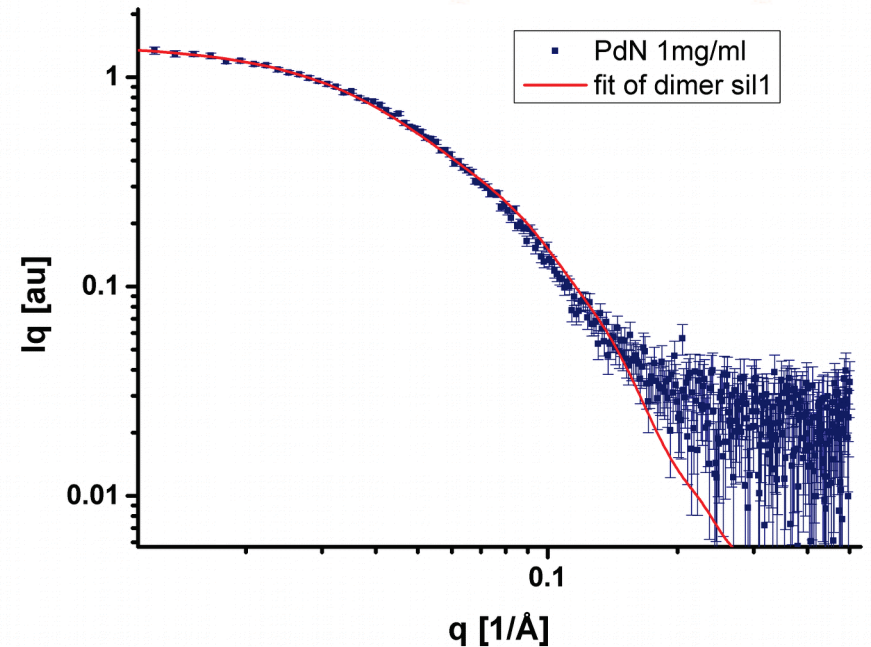
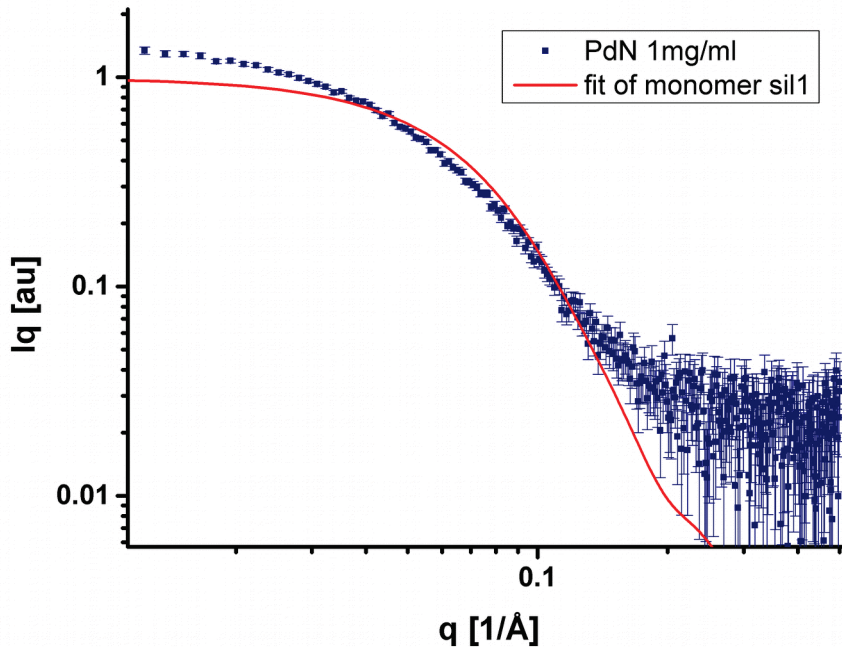
File	Dmax	Vp	MW
hBAPwt_b_1mg_as2_cnorm_a	200.3	136833.0	110.8
hBAPwt_b_1mg_as2_cnorm_h	171.3	161053.0	130.5
hBAPwt_b_2mg_as2_cnormf28_h	169.5	191366.0	155.0
hBAPwt_b_3mg_as2_cnorm-f1_a	134.2	157303.0	127.4
hBAPwt_b_3mg_as2_cnorm-f1_h	165.2	177899.0	144.1
hBAPwt_b_3mg_as2_cnorm-f48_a	164.4	232848.0	188.6
hBAPwt_b_3mg_as2_cnorm-f48_h	180.4	194074.0	157.2

hBAPwt 50 kD

## Fits

hBAPdN

Sil1



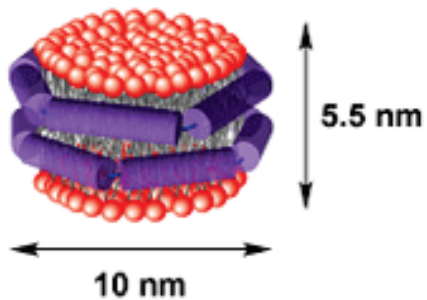
The monomer of Sil1 doesn't fit the scattering curve.

A dimer with dimerisation on the small edge fits the scattering much better.

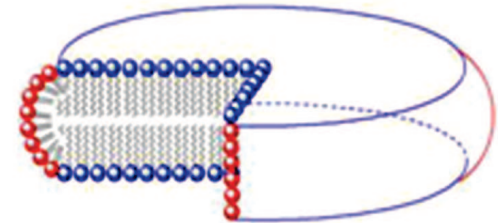
**The dN construct is a dimer, the N-terminus is not responsible for dimerisation.**

## System

Nanodisc



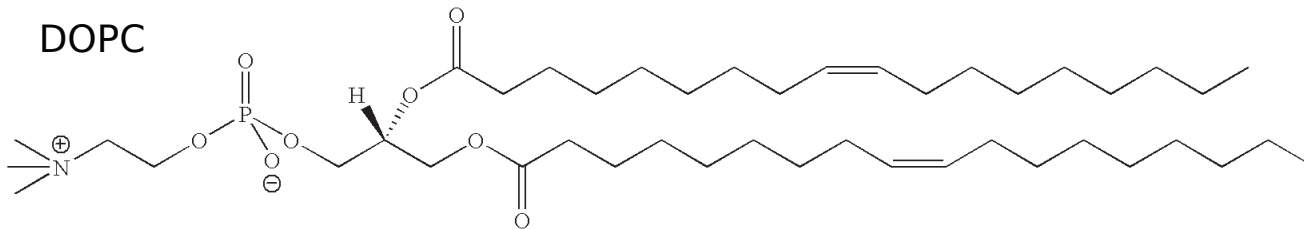
Model for fitting:  
Core Shell Bicelle



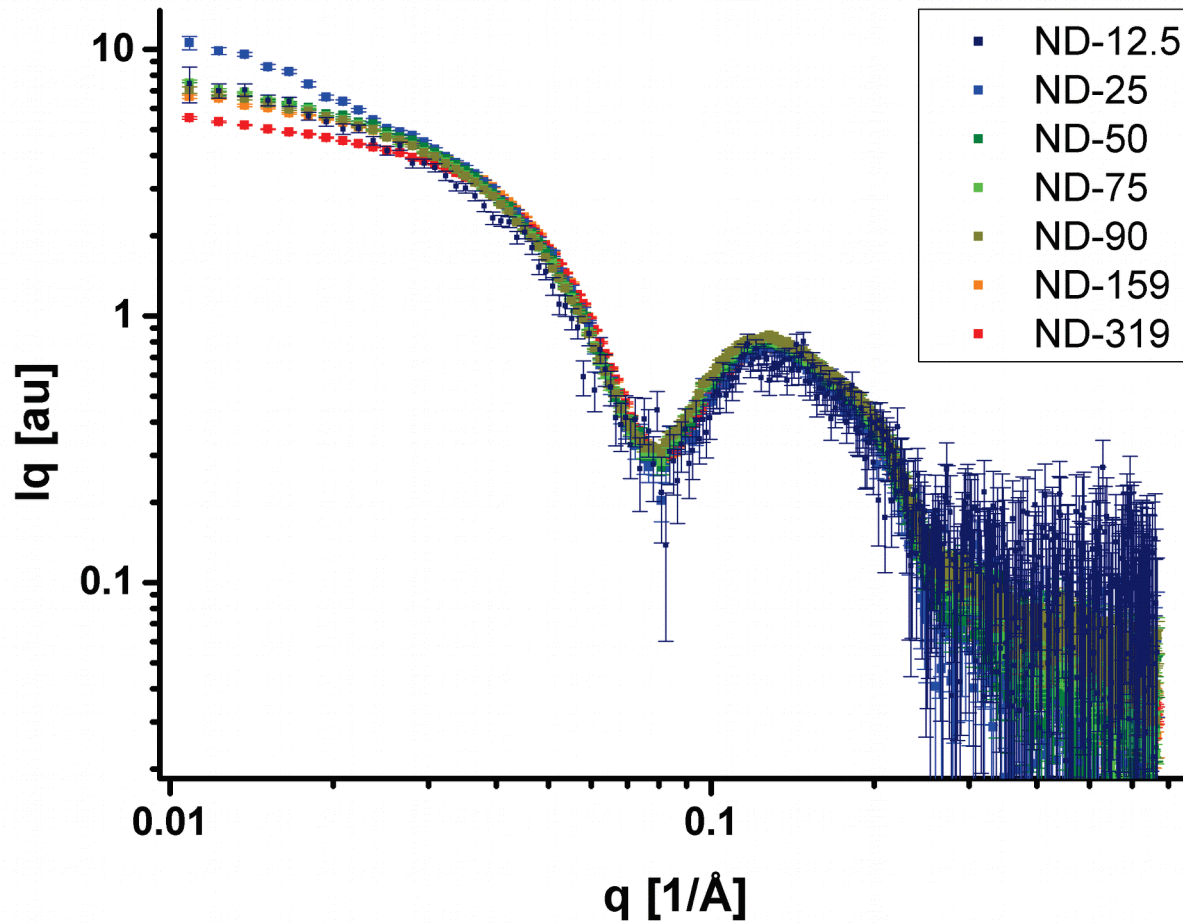
Bicelle

Membrane DOPC

Rim MSP ( $\alpha$ -helical, amphipathic membrane scaffold protein)

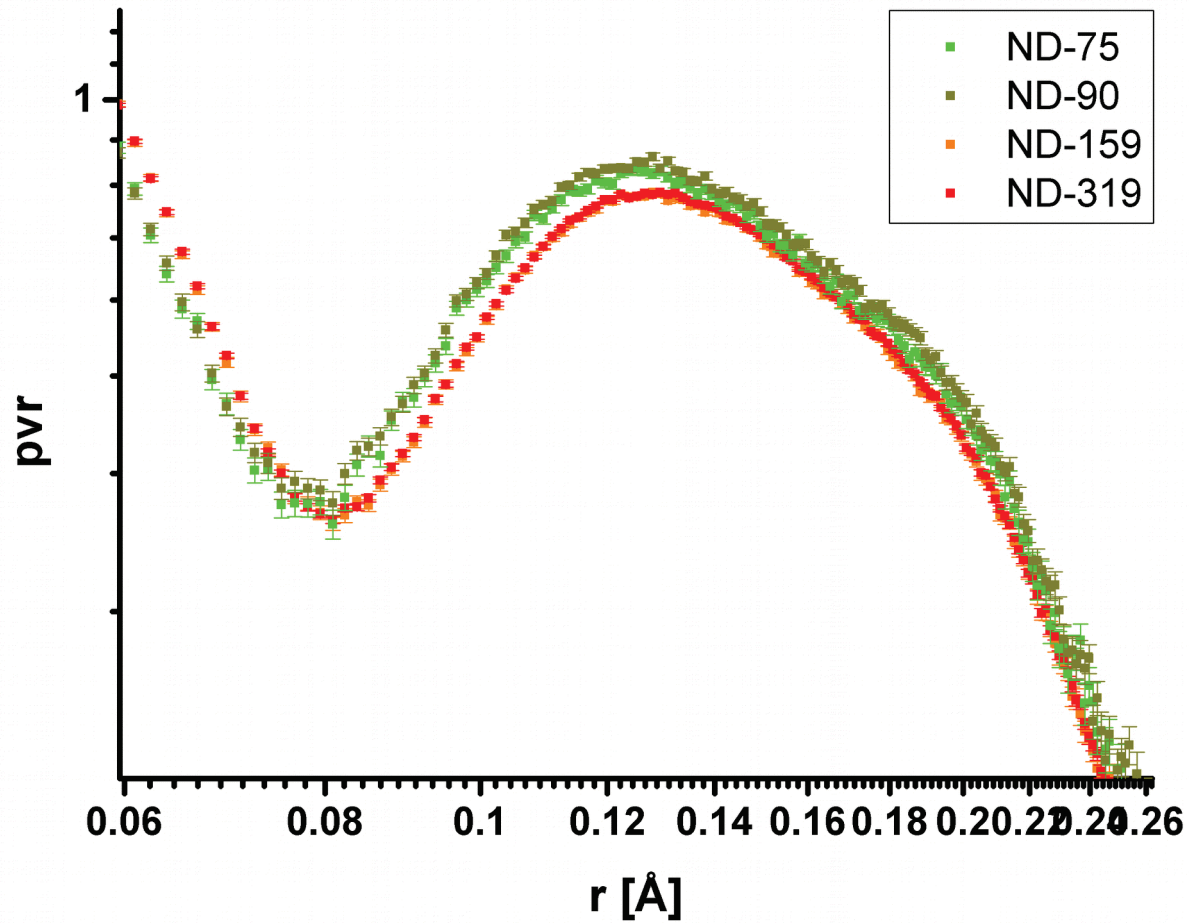


## Scattering



319 and 159  $\mu\text{mol}$  are affected by structure factor. 12.5  $\mu\text{mol}$  is too diluted and noisy

## Scattering



319 and 159  $\mu\text{mol}$  are not purified via sec minima are shifted.

## Fits



Model Parameters

<input checked="" type="checkbox"/> Select All	Value	Error	Min	Max	[Units]
<input checked="" type="checkbox"/> scale	13.657	+/- 2927.7	0	inf	
<input checked="" type="checkbox"/> background	0.060429	+/- 0.0008639	0	0.3	1/cm
<input checked="" type="checkbox"/> radius	10.004	+/- 0.24226	8	30	Ang
<input checked="" type="checkbox"/> rim_thickness	21.953	+/- 0.15844	12	25	Ang
<input checked="" type="checkbox"/> face_thickness	15.454	+/- 0.50046	3	20	Ang
<input checked="" type="checkbox"/> length	51.298	+/- 0.74995	30	60	Ang
<input checked="" type="checkbox"/> sld_core	8.4061	+/- 111.84	-inf	inf	1e-6/Ang <sup>2</sup>
<input checked="" type="checkbox"/> sld_face	10.277	+/- 88.707	-inf	inf	1e-6/Ang <sup>2</sup>
<input checked="" type="checkbox"/> sld_rim	9.5749	+/- 13.414	-inf	inf	1e-6/Ang <sup>2</sup>
<input checked="" type="checkbox"/> sld_solvent	9.3953	+/- 5.8309	-inf	inf	1e-6/Ang <sup>2</sup>

Polydispersity and Orientational Distribution

On  Off

Fitting

Set Instrumental Smearing

None  Use dQ Data  Custom Pinhole Smear  Custom Slit Smear

No smearing is selected...

Set Weighting by Selecting dI Source

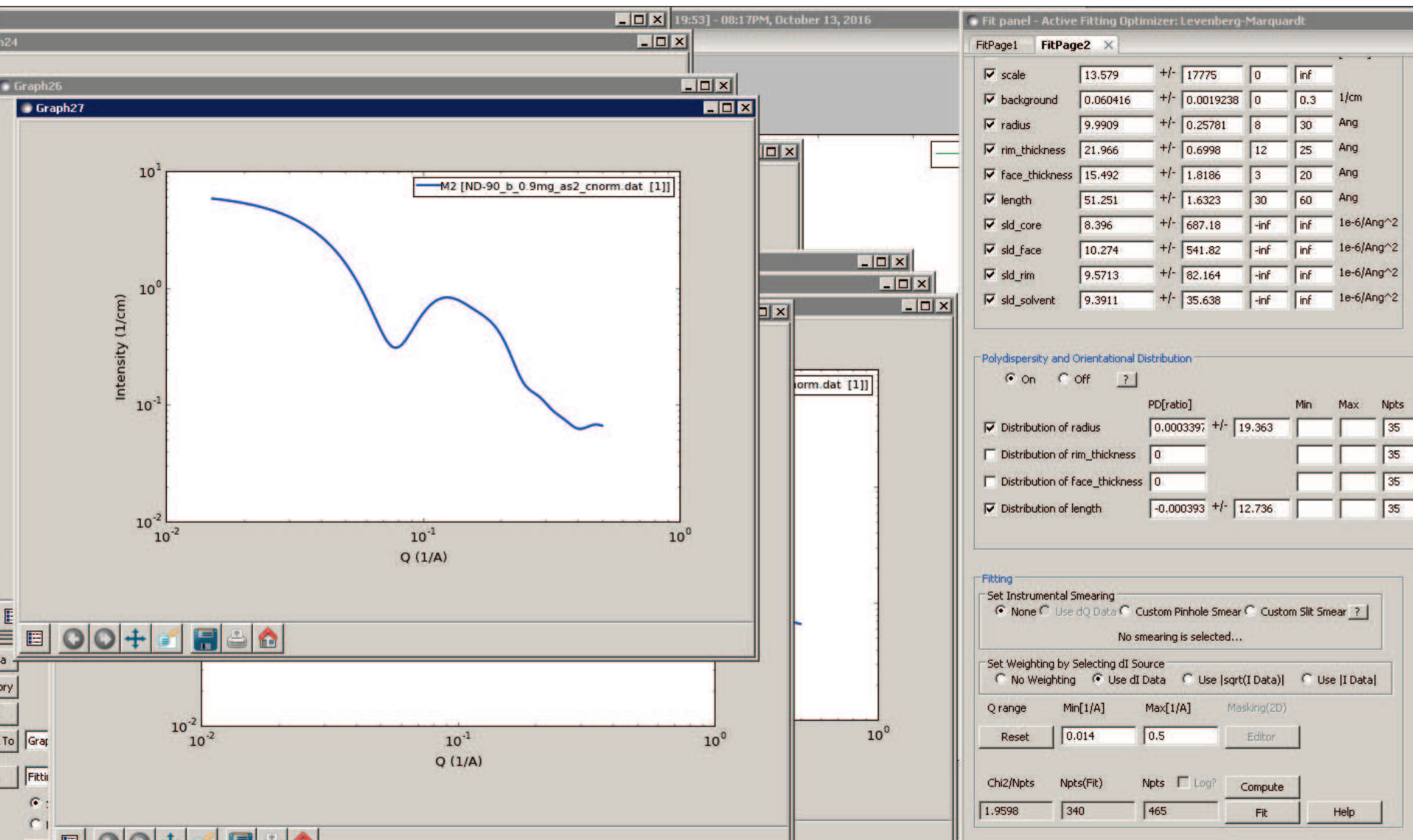
No Weighting  Use dI Data  Use |sqrt(I Data)|  Use |I Data|

Q range Min[1/A] Max[1/A] Masking(2D)

Reset

Chi2/Npts Npts(Fit) Npts  Log?

## Fits

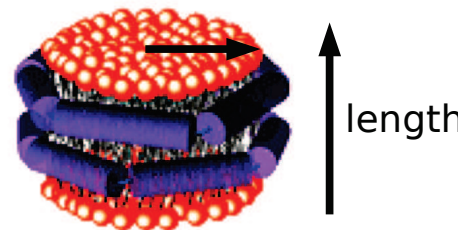


## Numbers

file	sc	bg	radius	rim_thickness	face_th	length	sld_core	sld_face	sld_rim	sld_solv	chi
ND319	8.62	0.04	10.25	19.817	17.869	43.46	8.3	10.3	9.69	9.46	15.94
ND159	9.9	0.04	10.044	21.046	16.537	47.426	8.3	10.3	9.62	9.4	1.45
ND159 PD	9.93	0.039	9.71 PD 0.11	21.433	15.62	48.745 PD -2.7e-6	8.25	10.32	9.61	9.39	1.36
ND90	9.79	0.06	9.903	22.11	15.669	51.232	8.25	10.477	9.658	9.44	-
ND90	13.657	0.0604	10.004	21.953	15.454	51.293	8.406	10.277	9.4749	9.39	1.94
ND90 PD	13.579	0.0604	9.9909 PD 0.000339	21.966	15.492	51.251 PD -0.00039	8.396	10.274	9.5713	9.3911	1.95
ND75	10.5	0.05	9.749	22.371	15.811	51.074	7.988	10.177	9.368	9.166	2.045
PD50	8.85	0.04	9.547	22.597	16.297	50.677	8.098	10.515	9.66	9.44	1.47
PD25	12.9	0.02	8.33	24.26	19.324	49.052	8.22	10.51	9.92	9.73	2.56
PD25 rdata	8.79	0.03	9.54	22.747	15.846	50.29	8.24	10.24	9.78	9.56	0.944
PD12p5	0.51	0.0012	10.358	22.366	15.674	53.776	8.00	10.377	9.66	9.37	1.16
PD12p5 rdata	0.51	0.0016	10.358	22.366	15.674	53.776	8.00	10.377	9.66	9.37	1.02

Poly dispersity  $PD = \delta r / r$

radius = radius of lipid layer



## Human HSP90

### **HSP90**

Monomer long axis 124 Å

Dimer open conformation long axis 175 Å

Should be in open conformation.

The human HSP90 binds TRb better in the open conformation.

Yeast HSP90 binds better in the closed conformation.

### **Thyroid hormone receptor (TRb-LBD)**

35 kD

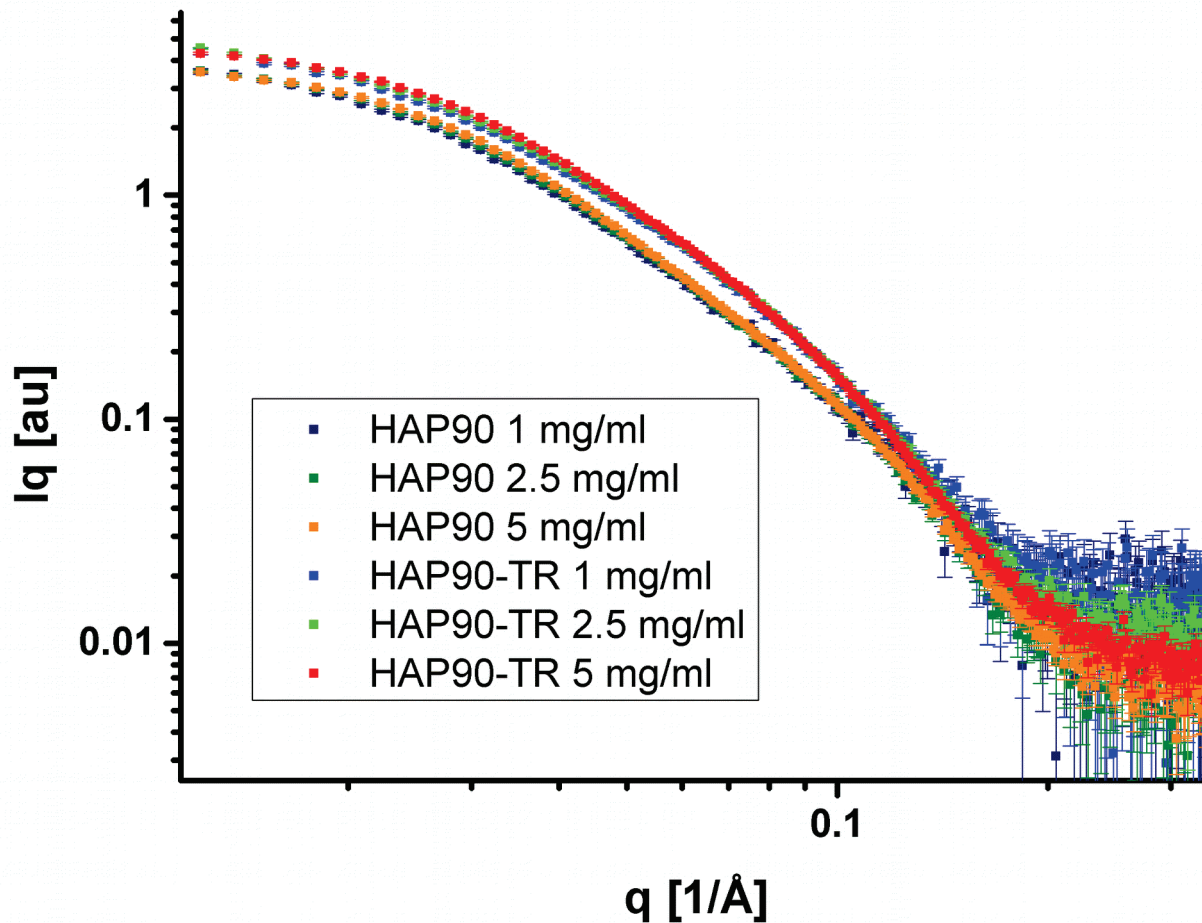
### **Questions:**

Is it binding?

Where?

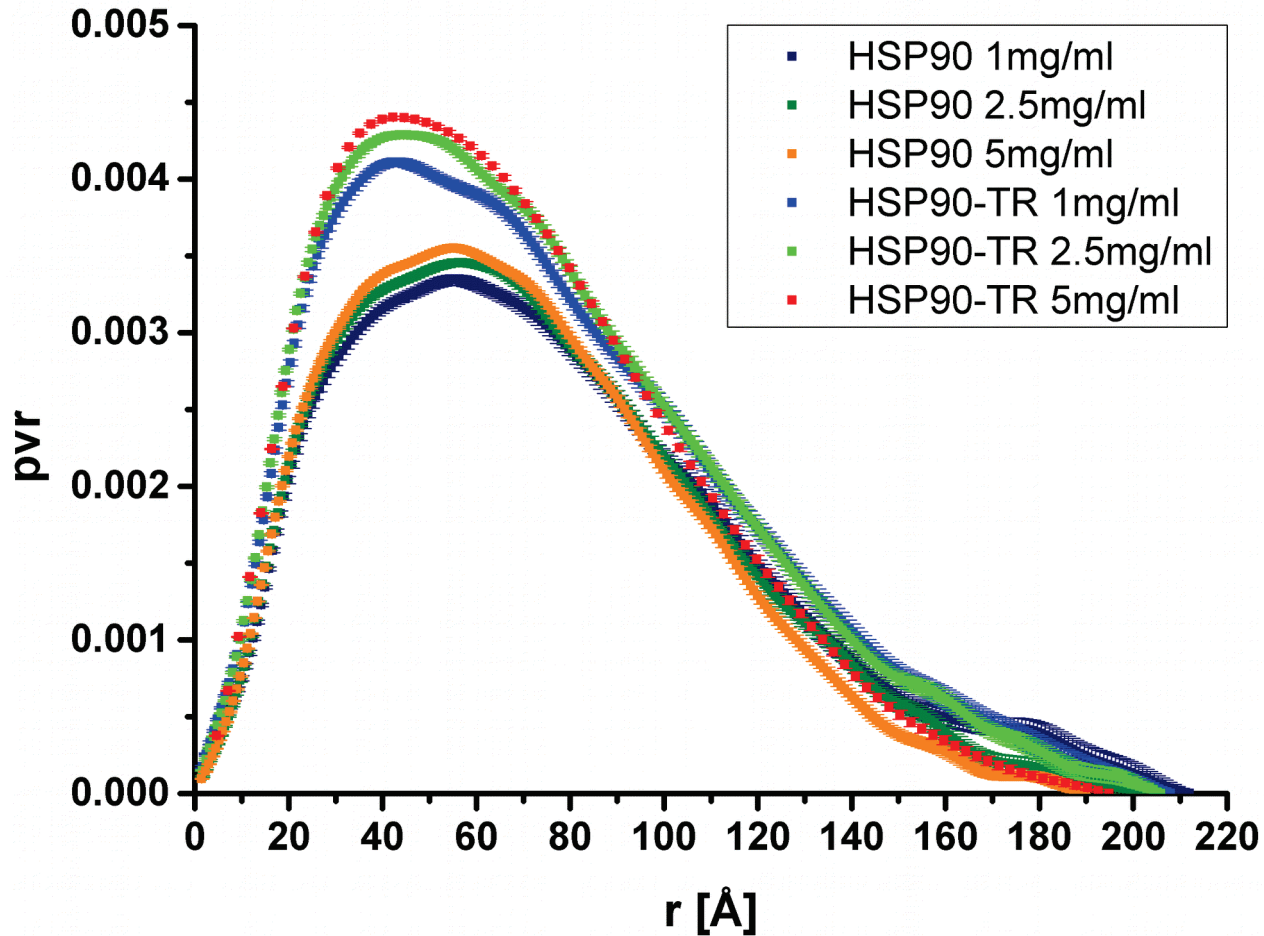
Stoichiometry?

## Scattering



Concentration dependent behaviour in the mid  $q$  region.

## Pair distance distribution



Dmax ~200 Å. peak shifted to smaller radius

## Numbers

File	Rg	+-	IO	+-	datarange	rgl
HSP90_a_1mg_as2_cnorm.dat	59.57	1.44			7-13	1.24
HSP90_a_2.5mg_as2_cnorm.dat	54.42	1			7-14	1.21
HSP90_a_5mg_as2_cnorm.dat	51.9	0.56			7-15	1.23
HSP90-TR_a_1mg_as2_cnorm.dat	56.74	1.15			7-14	1.27
HSP90-TR_a_2.5mg_as2_cnorm.dat	56.76	0.7			7-14	1.27
HSP90-TR_a_5mg_as2_cnorm.dat	50.9	0.24			7-16	1.28

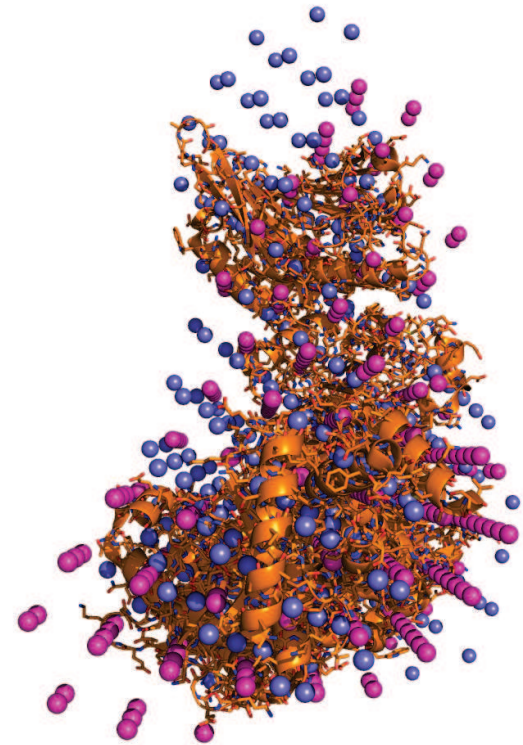
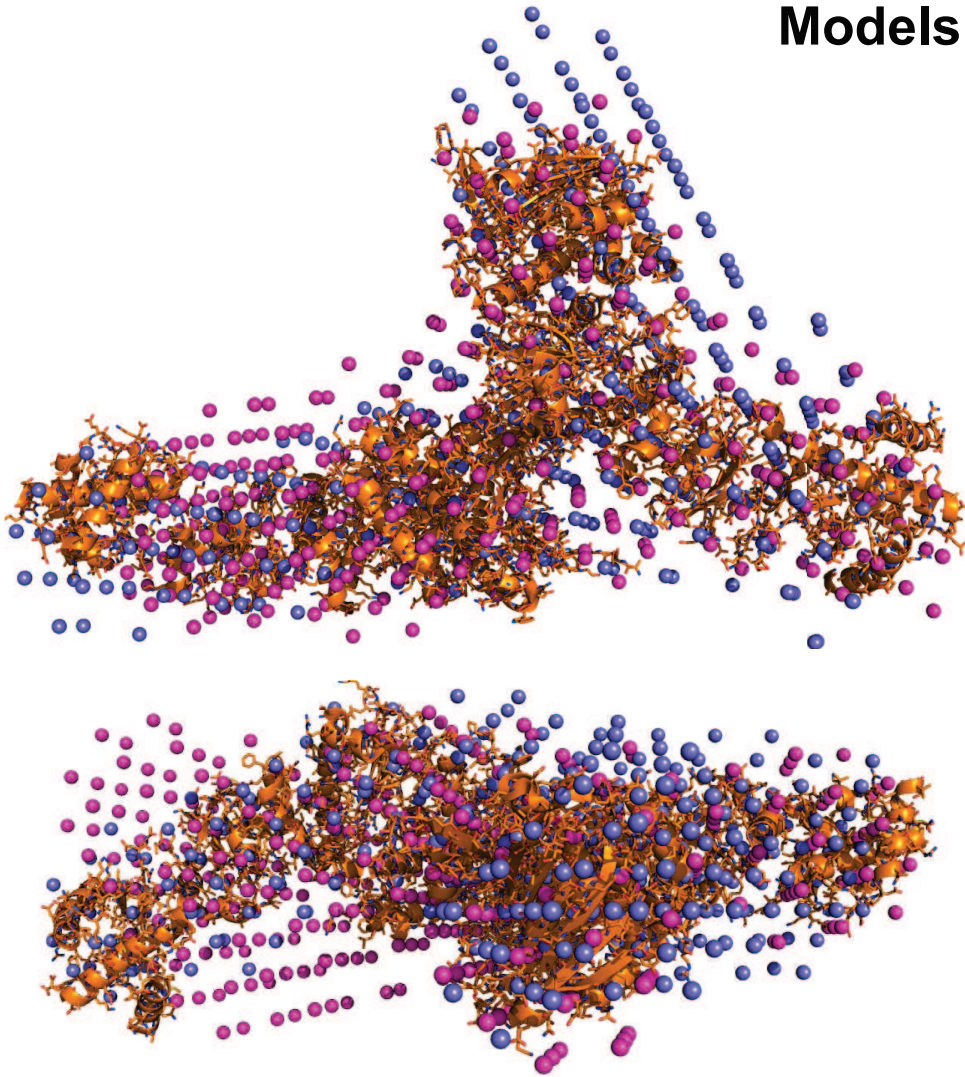
Concentration dependent shift in Rg

## Numbers

File	Dmax	Vp	MW
HSP90_a_1mg_as2_cnorm_h	212.0	432635.0	320.1
HSP90_a_2.5mg_as2_cnorm_h	205.0	395707.0	292.8
HSP90_a_5mg_as2_cnorm_h	190.0	361671.0	267.6
HSP90-TR_a_1mg_as2_cnorm_h	208.0	336122.0	248.7
HSP90-TR_a_2.5mg_as2_cnorm_h	206.0	330489.0	244.6
HSP90-TR_a_5mg_as2_cnorm_a	209.9	294178.0	217.7
HSP90-TR_a_5mg_as2_cnorm_h	194.9	298757.0	221.1

Concentration dependent shift in Dmax and MW

## Models



HSP90 5 mg/ml (magenta), HSP90-TR 5 mg/ml (blue)  
HSP90 open conformation (2IOQ)

Thanks

